PINEAPPLE JAM PHYSICOCHEMICAL AND SENSORY EVALUATION WITH ADDED PINEAPPLE PEEL

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ABSTRACT

The aim of this research was to evaluate the effect of pineapple peel (PP) addition on pineapple jam physicochemical and sensory characteristics. Five pineapple jam formulations were prepared: standard F1 (0% PP) and the others added 2.5% (F2), 5% (F3), 7.5% (F4) and 10% (F5) of PP. The results of sensory analysis indicated the feasibility of adding PP in pineapple jam, especially up to a level of 5%. The addition of PP increased soluble solids, titratable acidity, sugars, moisture, dietary fiber, phenolic compounds and antioxidant capacity. Also, it increased the values for a* and decreased for L* and b*. The use of up to 5% PP in jam should be encouraged as it can improve physicochemical characteristics and maintain sensory acceptability of the product. It also reduces negative effects of organic waste disposal on the environment.





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1. INTRODUCTION

Food waste is the food products disposal, which can occur from the beginning of the production chain to the final consumer [1]. On average 1.3 billion tons of food is wasted per year, resulting in approximately 30% of all food production. Roots, fruits, vegetables and oilseeds represent between 40 and 50% of these losses [2]. Brazil is among 10 countries that waste the most food in the world, close to 41 thousand tons. In the case of fruits, 30% of all production does not reach the final consumer's table [3]. After harvest, fruits have a very short shelf life, which leads to nutritional and economic losses. In addition, there is harm to the environment as the disposal of organic waste is increased. It is estimated that the rejection of byproducts from fruits and vegetables (seeds, stems, peel and leaves) can reach 30% in developed countries and 60% in developing countries [4]. However, it is known that it has a high nutritional content, especially bioactive compounds, which may help reduce the risk of cardiovascular disease, cancer and diabetes mellitus [5]. In this regard, there is a growing interest in using byproducts in food products, both by industry, researchers and consumers.

Pineapple (*Ananas comusus*) belongs to the Bromeliaceae family and, is one of the tropical fruits with the highest production in the world, estimated at around 20% [6]. Countries such as Thailand, the Philippines, Indonesia, India, China, Costa Rica, Brazil, Nigeria and South Africa are the largest fruit producers [7]. In 2017, world pineapple production was around 25 million tonnes, while in Brazil about 284,000 tons were produced, corresponding to approximately 11% of all fruit production [8]. In Brazil, most of pineapple production (97%) is destined for fresh consumption, being one of the six most consumed fruits [9]. This fruit has a good nutritional profile, with high levels of vitamins, minerals, fiber, flavonoids and carotenoids [10]. Generally, pineapple can also be consumed dehydrated, canned, as a juices, jams and compotes [7]. After harvesting fruit has a reduced shelf life, which leads to nutritional and economic losses. In addition, peel is also discarded despite having considerable antioxidants levels [11], [12], dietary fiber [12], vitamins and minerals [13]. However, the use of pineapple peel in food preparations as a form of nutritional enrichment has already shown positive results in jams [14], cereal bar [15] and yogurt [16]. The products showed good acceptability [14] and better technological and microbiological characteristics [15], [16]. Given this, the use viability of byproducts of pineapple is demonstrated, which can also contribute to environment preservation.

Commonly, jam is made from fruit pulp cooked, added water and sugar. Sometimes it may also contain citric acid and pectin [17]. Countries like Chile, China, United States, Spain, France, India, Turkey as well as Brazil stand out in jam production. In Brazil, production in 2017 was about 30.1 million tons of the product [18]. In 2016, 4,000 tons of jam were produced, with consumption reaching 3.36 billion tons [19]. Fruits such as grape, apricot, blueberry [20], mango, pineapple [21], strawberry, orange [17] and pomegranate [22] are the most used for jam production. As it is a low cost product and easy to process, storage and transport becomes a good alternative for the addition of unconventional ingredients such as fruit peels. Nevertheless, researches have already shown that replacing fruit pulp with peel in large quantities can reduce sensory acceptability, negatively alter firmness, color [22], elasticity, and final volume [23], in addition to nutritional characteristics of the product [24]. Considering this aspect, the aim of the present research was to evaluate the effect of pineapple peel (PP) addition in pineapple jam on the physicochemical and sensory characteristics.

2. MATERIALS AND METHODS

2.1. ETHICAL ISSUES

The study was conducted in accordance with the Declaration of Helsinki, and the protocol was approved by the Ethics Committee of Midwest State University, protocol number 2,201,325/2017.

2.2. RAW MATERIAL ACQUISITION

Six and a half kilograms of pineapple (*Ananas comosus*) was used for procedures. Fruits and ingredients were purchased from local shops in Guarapuava, PR, Brazil. Pineapples with better visual appearance were used. The peels were greenish yellow, without imperfections and of medium size (1.3 kg). All fruits were sanitized in running water and sanitized in sodium hypochlorite solution (150 ppm) for 10 minutes and again sanitized in running water. The

peel was extracted manually (0.5 cm), with the help of a knife, yielding 1.3 kg. The peel was crushed in a domestic blender (Mondial[®], Brazil) to form a homogeneous mass. It was then stored under refrigeration (8 °C) until the jams were prepared.

2.3. JAMS FORMULATIONS

Five pineapple jams formulations were prepared: standard F1 (0% PP) and the others added 2.5% (F2), 5% (F3), 7.5% (F4) and 10% (F5) of PP. These addition levels were defined by preliminary sensory testing performed with the product. In addition to PP, the following ingredients were used in formulations: pineapple pulp (65% (F1), 62.50% (F2), 60% (F3), 57.5% (F4) and 55% (F5)), sugar (34.4%), citric acid (Synth®, Brazil) (0.2%) and pectin (Mago®, Brazil) (0.4%). Fruit pulp and PP were mixed with sugar at room temperature (22 °C), stirring until sugar dissolved. The mixture was heated in domestic stove (Atlas®, Brazil) until boiling (103 °C). Then pectin was added and cooked for further 2 minutes. SS were measured and when they reached the ideal °Brix (63° to 68°) citric acid was added until pH was adjusted (3.30 to 3.40). Mixture was cooked for a further 2 minutes [25]. Jams were bottled still hot (79-85 °C) in 500 ml glass, pasteurized (70 °C) for 30 minutes, cooled to room temperature (22 °C) [26] and stored at room temperature (22 °C) until the analysis time.

2.4. SENSORY ANALYSIS

Sixty-three untrained volunteers participated in sensory analysis, being students, staff and teachers from UNICENTRO, of both genders, aged between 18 and 60 years. The tests were conducted in individual, white-lighted booths. Attributes of appearance, aroma, taste, texture and color and overall acceptance were assessed using a 9-point structured hedonic scale ranging from 1 ("very much disliked") to 9 ("very much liked"). Also, a purchase intention question was applied analyzed using a 5-point structured scale (1 - "would not buy" to 5 – "would buy for sure") [27]. Consumers received 10 g of each sample in disposable white plastic cups (50 mL), coded with three-digit numbers, in a randomized and balanced form, accompanied by a glass of water for palate cleansing. The formulations were offered in a monadic sequence. The Acceptability Index (AI) was calculated according to the formula: AI (%) = $A \times 100/B$ (A = mean grade obtained for the product; B = maximum grade given to the product) [28].

2.5. PHYSICOCHEMICAL ANALYSIS

Physicochemical determinations were performed in triplicate in pulp and peel, and in standard jam and in one with the highest percentage of peel and with sensory acceptance similar to standard product. Following evaluations were performed and results were expressed in wet matter: pH, measured by a bench pH meter (Tecnopon®, MPA-210 model, Brazil); SS, obtained by direct reading in ABBE bench refractometer (Bel®, RMI/RMT model, Brazil) [29]. Values were expressed in °Brix; TA using titration method [29]. Results were expressed as citric acid percentage (%); SS/TA ratio, which was obtained by dividing the values between SS and TA; Reducing Sugars (RS), Non-Reducing Sugars (NRS) and Total Sugars (TS) were evaluated by Lane-Eynon reductometric method [29]. Results were expressed as g 100 g⁻¹ and Color was analyzed by the Commission of the Commission Internationale de l'Eclairage (CIE) L^* (lightness), a^* (red-green) and b^* (yellow-blue), with colorimeter reading (Konica Minolta[®], Chroma Meter CR 4400 model, Tokyo, Japan) with illuminant D65 and angle 10° [30].

The moisture (g 100 g⁻¹), ash (g 100 g⁻¹) and protein (g 100 g⁻¹) contents were evaluated by the Association of Official Analytical Chemists [29] methodology; lipid (g 100 g⁻¹) by Bligh and Dyer [31]; carbohydrate (g 100 g⁻¹) by difference method (% Carbohydrate = 100 – (% moisture + % ash + % protein + % lipid + % fiber); total energy value (kcal 100 g⁻¹) using the values recommended by Atwater and Woods [32] for lipids (9 kcal g⁻¹), protein (4 kcal g⁻¹) and carbohydrate (4 kcal g⁻¹); total and insoluble dietary fiber (g 100 g⁻¹) determined by enzymatic method [29]. Soluble dietary fiber content (g 100 g⁻¹) was calculated by the difference in total and insoluble dietary fiber results; ascorbic acid (Vitamin C) determined by the 2.6 dichlorophenolindophenol titration method [29], modified by Benassi and Antunes [33] and results expressed in mg 100 g⁻¹; phenolic compounds measured by the Folin-Ciocalteu spectrophotometric method [34]. The values was taken on a spectrophotometer (Agillent Technologies[®], Cary 60 UV model, Malaysia) at 765 nm and results being expressed as mg of gallic acid equivalent (GAE) 100 g⁻¹; total carotenoids (µg g⁻¹) obtained by spectrophotometric analysis (Agillent Technologies[®], Cary 60 UV model, Malaysia)

at 450 nm [35]; antioxidant capacity was evaluated by the ABTS method (2,2-azinobis-[3-ethyl-benzothiazolin-6-sulfonic acid]) in both versions hydrophilic with ABTS solution was diluted with acetone 50% and lipophilic with ABTS solution was diluted with methanol 50% [36]. Results were expressed in µmol Trolox 100⁻¹ of sample.

2.6. STATISTICAL ANALYSIS

Data were analyzed using software R version 3.6.1 by analysis of variance (ANOVA). The comparison of means was performed by Tukey and Student's t-tests, evaluated with a 5% significance level.

3. RESULTS AND DISCUSSIONS

3.1. SENSORY ANALYSIS

Table 1 describes the sensory scores of jams added with different PP levels. Higher scores for appearance (p < 0.05) were found to F1, F2 and F3 formulations. The addition of PP in jams did not affect aroma notes (p > 0.05). Higher acceptability for taste, texture and color attributes was observed for formulations F1 and F2, compared to F4 and F5. Also, F3 was better accepted than F5. Samples F1 and F2 scored higher than F4 and F5 considering overall acceptance, while F3, F4 and F5 did not differ significantly (p > 0.05). Purchase intention scores were lower for F4 and F5 formulations (p < 0.05). Thus, it can be seen that the addition of up to 5% PP in pineapple jam does not interfere with sensory acceptability, corroborating the Younis et al. [24] research that investigated papaya jam with sweet lemon peel flour (5%). The presence of high levels of phenolic compounds in PP (Table 2) may explain the lower grades obtained in samples F4 and F5. When heated, these substances cause an astringency sense in product [37] which impairs acceptability.

contents								
Parameter	0%	2.5%	5%	7.5%	10%			
Appearance	8.0 ± 1.43^{a}	7.7 ± 1.28^{a}	7.2 ± 1.20^{a}	6.3±2.05 ^b	5.8±1.94 ^b			
AI (%)	88.3	85.0	80.6	70.3	64.3			
Aroma	7.6 ± 1.27^{a}	7.5 ± 1.42^{a}	7.5 ± 1.39^{a}	7.0 ± 1.64^{a}	6.9 ± 1.66^{a}			
AI (%)	84.8	83.1	83.2	77.8	77.1			
Taste	7.7 ± 1.41^{a}	7.3 ± 1.31^{a}	7.0 ± 1.49^{ab}	6.6±1.93 ^{bc}	6.1±1.99 ^c			
AI (%)	85.6	81.6	77.8	73.8	67.2			
Texture	7.3±1.81 ^a	6.6 ± 1.88^{a}	6.3±1.29 ^{ab}	5.4 ± 2.11^{bc}	4.7±2.33 ^c			
AI (%)	80.9	73.7	69.6	59.7	51.9			
Color	7.8 ± 1.53^{a}	7.5 ± 1.53^{a}	7.0 ± 1.37^{ab}	6.5±1.83 ^{bc}	5.8±2.06 ^c			
AI (%)	87.1	83.4	78.0	72.1	64.9			
Overall Acceptance	7.1±1.99 ^a	7.1±1.65ª	6.2±1.69 ^{ab}	6.1±2.13 ^b	5.4±2.11 ^b			
AI (%)	78.9	78.7	69.4	67.9	60.1			
Purchase Intention	4.0 ± 1.17^{a}	3.7 ± 1.06^{ab}	3.4±0.90 ^{ab}	3.2±1.34 ^b	2.7±1.19°			

Table 1: Sensory scores (mean ± standard deviation) of pineapple jam added with different pineapple peel

contonto

Distinct letters on the same row indicate significant difference in the Tukey's test (p < 0.05) for jams: AI: Acceptability Index.

Color is an attribute directly related to food product acceptance. During the jam processing, it was observed that the greater was the addition of peel, the darker was the color. Carotenoid instability at high temperatures [38] may explain this effect. Carotenoids are highly unsaturated compounds and are subject to oxidation and isomeration reactions that decrease biological activity and alter the characteristic color of these compounds [39]. The reaction is favored by the presence of light, heat, metals, enzymes and peroxides [40]. Addition of PP altered the jam texture as it made gel formation difficult, promoting more pasty consistency and formation of fibrous residues in product. In jams made only with fruit pulp, gel formation occurs due to the balance between pectin, sugar and acid contents. With the peel addition, this process is compromised [17], as it contains higher fiber contents than pulp [41], International Journal of Research -GRANTHAALAYAH

increasing liquid absorption and, consequently, product consistency [42]. In general, peel addition up to 5% in pineapple jam maintained an AI \ge 70%, which ranks the product with good sensory acceptance [43]. This generally indicates the feasibility of adding PP to the jam, especially up to a level of 5%. Similar results were observed after the use of PP in pineapple jam [44] and PP extract in peach jam [14].

3.2. PHYSICOCHEMICAL COMPOSITION

Considering that sample F3 was the one with the highest PP content and with similar sensory acceptance as standard in all evaluations (Table 1), it was selected for physicochemical comparison purposes along with the standard formulation (F1) presented in Table 2. Higher pH levels were verified for PP while the pulp presented higher Titratable Acidity (TA). Nevertheless, PP addition reduced pH and increased TA in jam (5% PP), as verified by Lago-Vanzela et al. [45] evaluating jam with addition of 'cajá-manga' peel. The peel has higher amounts of polysaccharides in the form of pectin, cellulose and hemicellulose [46]. In acidic solutions, these substances are depolymerized, generating compounds such as xylose, acetic acid, furfural and hydroxymethylfurfural which can be transformed into formic acid [47], increasing the acidity of the product.

Parameter	Pineapple	Pineapple	F1	F3
	pulp	peel		
рН	3.8±0.01 ^B	6.3±0.01 ^A	3.4±0.01 ^a	3.3±0.02 ^b
Soluble Solids (^o Brix)	20.0±0.00 ^A	2.1 ± 0.00^{B}	63.0±0.00 ^b	65.0±0.00ª
Titratable Acidity (% citric acid)	$7.0 \pm 0.04^{\text{A}}$	0.6 ± 0.02^{B}	1.1 ± 0.04^{b}	1.3 ± 0.02^{a}
Soluble Solids/Titratable Acidity (ratio)	2.9 ± 0.02^{B}	$3.3 \pm 0.08^{\text{A}}$	50.6 ± 0.02^{a}	51.1 ± 0.02^{a}
Reducing Sugars (g 100 g ⁻¹)	4.6±0.03 ^A	3.0 ± 0.02^{B}	14.3 ± 0.00^{b}	25.0 ± 0.01^{a}
Nonreducting Sugars (g 100 g ⁻¹)	12.1 ± 0.01^{B}	15.3±0.26 ^A	21.4 ± 0.01^{b}	27.6 ± 0.02^{a}
Total Sugars (g 100 g ⁻¹)	16.7 ± 0.00^{B}	$18.3 \pm 0.0^{\text{A}}$	35.7±0.01 ^b	52.6±0.01 ^a
L*	64.7±0.59 ^A	51.2±0.77 ^B	32.9±0.83 ^a	29.7 ± 0.78^{b}
a*	-4.5 ± 0.08^{B}	-2.7±0.43 ^A	-1.6 ± 0.10^{b}	-1.1±0.11 ^a
b*	18.0 ± 0.53^{B}	$20.7 \pm 0.82^{\text{A}}$	6.6 ± 0.25^{a}	4.6 ± 0.34^{b}
Moisture (g 100 g ⁻¹)	85.4±0.06 ^A	84.9 ± 0.05^{B}	20.9 ± 0.03^{b}	32.7 ± 0.07^{a}
Ash (g 100 g-1)	0.3 ± 0.03^{B}	$0.7 \pm 0.02^{\text{A}}$	0.3 ± 0.07^{a}	0.3 ± 0.08^{a}
Protein (g 100 g ⁻¹)	0.7 ± 0.05^{B}	$0.8 \pm 0.04^{\text{A}}$	0.3 ± 0.06^{a}	0.4 ± 0.09^{a}
Lipid (g 100 g ⁻¹)	0.0 ± 0.06^{B}	$0.1 \pm 0.04^{\text{A}}$	0.1 ± 0.09^{a}	0.1 ± 0.04^{a}
Carbohydrate (g 100 g ⁻¹)*	13.6±0.45 ^A	13.6±0.86 ^A	78.4 ± 0.56^{a}	66.6±0.23 ^b
Total energy value (kcal 100 g ⁻¹)	57.3±0.69 ^A	58.1±0.98 ^A	315.8 ± 0.64^{a}	268.3±0.76 ^b
Soluble fiber (g 100 g ⁻¹)**	0.0 ± 0.11^{B}	$0.3 \pm 0.09^{\text{A}}$	0.1 ± 0.15^{a}	0.1 ± 0.11^{a}
Insoluble fiber (g 100 g ⁻¹)**	0.8 ± 0.13^{B}	$5.1 \pm 0.10^{\text{A}}$	0.5 ± 0.10^{b}	0.7 ± 0.12^{a}
Total fiber (g 100 g ⁻¹)**	0.8 ± 0.10^{B}	5.4 ± 0.14^{A}	0.6 ± 0.16^{b}	0.8 ± 0.15^{a}
Total carotenoid (μg g ⁻¹)	24.9±0.03 ^A	10.9 ± 0.01^{B}	3.1 ± 0.06^{a}	2.9±0.01 ^b
Ascorbic acid (mg 100 g ⁻¹)	120.8±0.02 ^A	6.3±0.06 ^B	13.5 ± 0.12^{a}	12.8 ± 0.48^{b}
Phenolic compounds (mg GAE 100 g ⁻¹)	5.6±0.03 ^B	39.8±0.03 ^A	35.9±0.01 ^b	40.5 ± 0.03^{a}

Table 2: Physicochemical composition (mean \pm standard deviation) of pineapple pulp, peel and jam with added 0% (F1) and 5% (F3)

Distinct letters in row are significantly different by the Student's t-test (p < 0.05), considering capital letters for the differences between pulp and peel and lowercase letters for the jams; Results reported in wet weight basis; *not include dietary fiber; **dietary fiber.

Soluble Solids (SS) content together with TA (SS/TA) is one of the most efficient methods for demonstrating the degree of fruit maturity and taste. Generally, products with higher SS content and lower TA present greater sweetness and acceptability [48]. Although pineapple pulp had higher SS content and lower SS/TA ratio than peel (p < 0.05), the jam containing PP had higher SS concentration and SS/TA ratio similar to F1. During cooking, polysaccharides hydrolysis (pectin, cellulose and hemicellulose) occurs [49] present in peel, which results in glucose

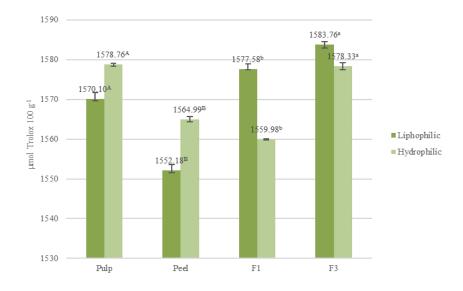
and fructose formation [50]. In addition, pectin is present in higher concentration in peel (2.49 g 100 g⁻¹) [49] compared to pulp (91.08 mg 100 g⁻¹) [51]. This fiber has hydrophilic groups that absorb water, which concentrates SS [52], corroborating to literature [45].

Pulp presented higher Reducing Sugars (RS) (glucose and fructose) and lower Nonreducting Sugars (NRS) (sucrose) and Total Sugars (TS) contents in relation to peel. In peel, the RS are responsible for the fruit protection [53]. During ripening, starch is converted into glucose and fructose, which are used in stressful situations, such as at low temperatures, for example, which explains the lower RS content in peel [53]. The peel addition to jam increased RS, NRS and TS concentration. Cooking in an acid medium promotes sucrose degradation, present in higher amounts in peel, in glucose and fructose [54], which increases the RS concentration in jam.

Higher values for L* and lower than a* and b* were found in pineapple pulp when compared to peel. Pulp and peel a* values were negative, which demonstrates a greenish color. Similar results were observed in pineapples grown in Thailand [55]. In the case of pineapple, one of the most common methods for analyzing ripeness is the changing of the peel color from green to yellow [10]. In jam, the addition of PP increased the value of a* and decreased the values of L* and b*, as it has lower brightness and higher red content than pulp. This is due to the presence of higher chlorophyll content present in pineapple peel (232.13 μ g g⁻¹) compared to pulp (152.12 μ g g⁻¹) [56], since when degraded it has a greenish brown color [57]. In general, jams can be considered dark in color, as L* values were less than 50%, with a yellow tone (b*) and a green sub-tonality (a*).

Pulp had higher moisture content compared to peel, since it has high water content (85-86%) [10]. However ash, protein, lipid and fiber contents were higher for the peel, as these compounds have protective function, preventing oxidation [58], mechanical damage [59] and action of microorganisms [60]. There was no significant difference (p > 0.05) between carbohydrate and calorie contents between pulp and peel. The addition of PP increased moisture content in jam (p < 0.05) due to its higher fiber content, which has high hygroscopic content [57]. There was no significant difference (p > 0.05) between jams for ash, protein and lipid contents. However, the PP use in jam reduced carbohydrate and calorie content and increased fiber content as found by Damiani et al. [61] in jelly with mango peel addition (0 to 100%).

Pineapple pulp presented higher levels of total carotenoids and ascorbic acid which consequently reduced the content of these compounds in F3 (p < 0.05). Although pineapple pulp had approximately 19 times more vitamin C than the peel, the difference in vitamin content between F1 and F3 was small. Compounds such as flavonoids, present in larger quantities in peel (211.20 mg QE 100 g⁻¹) [62] than in pulp (26.20 mg GAE 100 g⁻¹) [63], prevent the vitamin C degradation, as they are potent antioxidants. Pineapple peel presented higher levels of phenolic compounds than pulp, which significantly increased phenolic content in jam.



Distinct letters in column are significantly different by the Student's t-test (p < 0.05), considering capital letters for the differences between pulp and peel and lowercase letters for the jams; Results reported in wet weight basis.

Figure 1: Lipophilic and hydrophilic mean antioxidant capacity of pineapple pulp, peel, and jam with added 0% (F1) and 5% (F3).

Figure 1 shows results of the antioxidant capacity for pulp and peel pineapple and standard jam (F1) and added with 5% (F3) of PP. Higher antioxidant capacity was observed in pulp compared to PP (p < 0.05), since the pulp has higher amounts of ascorbic acid and carotenoids than the peel. However, PP addition increased antioxidant capacity in jam, similar to that observed by Damiani et al. [61]. This effect occurs due to the high cooking temperature, promoting the hydrolysis of vacuoles [49] and lipid membranes [64] present in peel cell wall. This results in release of a significant content of phenolic compounds [65] and carotenoids [64] in jam.

4. CONCLUSIONS

A level up to 5% addition of pineapple peel in jam is well accepted by consumers, achieving sensory acceptance similar to standard product. Moreover, it improves physicochemical profile of the product with increased antioxidant capacity and soluble solids, titratable acidity, sugars, fiber and phenolic compounds, with reduced carbohydrate and calorie content. The use of jam byproducts should be encouraged as it can improve their physicochemical characteristics and maintain sensory acceptability. It also reduces the negative effects of organic waste disposal on the environment.

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CONFLICT OF INTEREST

The author have declared that no competing interests exist.

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REFERENCES

- [1] Food and Agriculture Organization of the United Nations (FAO). Food Wastage Footprint: Impacts on Natural Resources: Summary Report. Rome: FAO; 2013.
- [2] Food and Agriculture Organization of the United Nations (FAO). Food losses and waste in latin American and the Carribean. Rome: FAO; 2014.
- [3] Food and Agriculture Organization of the United Nations (FAO). FAO apresenta avanços no combate às perdas e ao desperdício de alimentos. Available online: http://www.fao.org/brasil/noticias/detailevents/en/c/1062706/ (accessed on 12 May 2020).
- [4] Hoornweg D, Bhada-Tata P. What a Waste: A Global Review of Solid Waste Management. Washington: World Bank; 2012.
- [5] Zhang YJ, Gan RY, Li S, Zhou Y, Li AN, Xu DP, Li HB. Antioxidant phytochemicals for the prevention and treatment of chronic diseases. Molecules. 2015;20(12):21138-21156.
- [6] Lu XH, Sun DQ, Wu QS, Liu SH, Sun, GM. Physico-chemical properties, antioxidant activity and mineral content of pineapple genotypes grown in China. Molecules. 2014;19(6):8518–8532.
- [7] Lobo MG, Paull RE. Handbook of pineapple technology: production, postharvest science, processing and nutrition. Chichester: John Wiley & Sons; 2017.
- [8] Food and Agriculture Organization of the United Nations (FAO). Major Tropical Fruits Market Review 2017. Rome: FAO; 2019.

- Instituto Brasileiro de Geografia e Estatística (IBGE). Produção agrícola municipal. Rio de Janeiro: IBGE; 2013. [9]
- [10] Ancos B, Sánchez-Moreno C, González-Aguilar GA. Pineapple composition and nutrition. In: Ancos B, Moreno CS, Aguilar GAG. Handbook of pineapple technology: production, postharvest science, processing and nutrition. Chichester: John Wiley & Sons; 2017.
- [11] Silva AC, Jorge N. Bioactive compounds of the lipid fractions of agro-industrial waste. Food Research International. 2014;66(1):4939-500.
- [12] Morais DR, Rotta EM, Sargi SC, Schmidt EM, Bonafe EG, Eberlin MN, Sawayad ACHF, Visentainer JV. Antioxidant activity, phenolics and UPLC-ESI (-)-MS of extracts from different tropical fruits parts and processed peels. Food Research International. 2015;77(part 3):392-399.
- [13] Van der Goot AJ, Pelgrom PJ, Berghout JA, Geerts ME, Jankowiak L, Hardt NA, Keijer J, Schutyser MAI, Nikiforidis CV, Boom RM. Concepts for further sustainable production of foods. Journal of Food Engineering. 2016;168(1):42-51.
- [14] Vieira ECS, Silva EP, Amorim CCM, de Sousa GM, Becker FS, Damiani C. Aceitabilidade e características físicoquímicas de geleia mista de casca de abacaxi e polpa de pêssego. Científica. 2017;45(2):115-122.
- [15] Damasceno AK, Gonçalves CAA, Pereira GS, Costa LL, Campagnol PCB, Almeida PL, Arantes-Pereira L. Development of cereal bars containing pineapple peel flour (Ananas comosus L. Merril). Journal of Food Quality. 2016;39(5):417-424.
- [16] Sah BNP, Vasiljevic T, McKechnie S, Donkor ON. Physicochemical, textural and rheological properties of probiotic yogurt fortified with fibre-rich pineapple peel powder during refrigerated storage. LWT-Food Science and Technology. 2016;65(1):978-986.
- [17] Featherstone SA. Complete Course in Canning and Related Processes: Processing Procedures for Canned Food Products. 14nd ed. Waltham: Elsevier; 2015.
- [18] Associação Brasileira das Indústrias de Alimentação (ABIA). Produção e consumo de geleias. Available online: https://www.abia.org.br/ (accessed on 15 May 2020).
- [19] Centre for the Promotion of Imports from Developing Countries (CBI). Exporting fruit juices to Europe. Available online: https://www.cbi.eu/market-information/processed-fruit-vegetables-edible-nuts/fruitjuices/europe/ (accessed on 03 May 2020).
- [20] Naeem MM, Fairulnizal MM, Norhayati MK, Zaiton A, Norliza AH, Syuriahti WW, Azerulazree JM, Aswir AR, Rusidah S. The nutritional composition of fruit jams in the Malaysian market. Journal of the Saudi Society of Agricultural Sciences. 2017;16(1):89-96.
- [21] Asema SUK, Parveen N. Study of Heavy Metal Content by AAS in a Variety of Flavours of Jam Samples and Its Physicochemical Characterization. International Journal of Scientific Research in Science, Engineering and Technology. 2018;4(1):1259-1261.
- [22] Abid M, Yaich H, Hidouri H, Attia H, Ayadi MA. Effect of Substituted Gelling Agents from Pomegranate Peel on Colour, Textural and Sensory Properties of Pomegranate Jam. Food Chemistry. 2018;239(1):1047-1054.
- [23] Fu JT, Shiau SY, Chang RC. Effect of calamondin fiber on rheological, antioxidative and sensory properties of dough and steamed bread. Journal of Texture Studies. 2014;45(5):367-376.
- [24] Younis, K, Islam, R.U, Jahan, K, Yousuf, B, Ray, A. Effect of Addition of Mosambi (Citrus Limetta) Peel Powder on Textural and Sensory Properties of Papaya Jam. Cogent Food & Agriculture. 2015;1(1):1-8.
- [25] Phillips GO, Williams PA. Handbook of Hydrocolloids. 2nd ed. Boca Raton: Woodhead Publishing; 2009.
- [26] Pereira AGT, Pereira PAP, Borges SV, Dias MV, Figueiredo LP, Valente WA. Physicochemical characterization and sensory evaluation of jellies made with guava peels (Psidium guajava L.). International Journal of Agricultural Policy and Research. 2015;3(11):396-401.
- [27] Meilgaard M, Civille GV, Carr BT. Sensory Evaluation Techniques. 5nd ed. Boca Raton: CRC Press; 2016.
- [28] Macfie HJ, Bratchell N, Greenhof K, Vallis LV. Designs to balance the effect of order of presentation and firstorder carry over effects in hall tests. Journal of Sensory Studies. 1989;4(2):129-148.
- [29] Association of Official Agricultural Chemists (AOAC) International. Official Methods of Analysis of AOAC International. Gaithersburg: Association of Official Analytical Chemists; 2011.
- [30] Konica Minolta. Precise Color Communication: Color Control from Perception to Instrumentation; Ramsey: Konica Minolta; 2007.
- [31] Bligh EG, Dyer WJ. A Rapid Method of Total Lipid Extraction and Purification. Canadian Journal of Biochemistry and Physiology. 1959;37(8):911-917. International Journal of Research -GRANTHAALAYAH

- [32] Atwater WO, Woods CD. The chemical composition of American food materials. US Official Experiment Stations. Experiment Station Bulletin. 1896;28(1):461-462.
- [33] Benassi MT Antunes AJ. A Comparison of Metaphosphoric and Oxalic Acids as Extractants Solutions for the Determination of Vitamin C in Selected Vegetables. Biology and Technology Archives. 1988;31(4):507-513.
- [34] Bucić-Kojić A, Planinić M, Tomas S, Bilić M, Velić D. Study of Solid–Liquid Extraction Kinetics of Total Polyphenols from Grape Seeds. Journal of Food Engineering. 2007;81(1):236-242.
- [35] Rodriguez-Amaya DB, Kimura M. Handbook for Carotenoid Analysis; International Food Policy Research Institute (IFPRI) and International Center for Tropical Agriculture (CIAT): Washington: HarvestPlus; 2004.
- [36] Miller NJ, Sampson J, Candeias LP, Bramley PM, Rice-Evans CA. Antioxidant Activities of Carotenes and Xanthophylls. FEBS Letters. 1996;384(3):240-242.
- [37] Mäkilä L, Laaksonen O, Kallio H, Yang B. Effect of processing technologies and storage conditions on stability of black currant juices with special focus on phenolic compounds and sensory properties. Food Chemistry. 2017;221(1):422-430.
- [38] Maiani G, Castón MJ, Catasta G, Toti E, Cambrodón IG, Bysted A, Granado-Lorencio F, Olmedilla-Alonso B, Knuthsen P, Valoti M, Böhm V, Mayer-Miebach E, Behsnilian D, Schlemmer, U. Carotenoids: actual knowledge on food sources, intakes, stability and bioavailability and their protective role in humans. Molecular Nutrition & Food Research. 2009;53(S2): S194-S218.
- [39] Rodriguez-Amaya DB. Food carotenoids: analysis, composition and alterations during storage and processing of foods. Forum of Nutrition. 2003;56(1):35-37.
- [40] Alves RMV, Ito D, Carvalho JLV, Godoy RDO. Estabilidade de farinha de batata-doce biofortificada. Brazilian Journal of Food Technology. 2012;15(1):59-71.
- [41] Martínez R, Torres P, Meneses MA, Figueroa JG, Pérez-Álvarez JA, Viuda-Martos M. Chemical, technological and in vitro antioxidant properties of mango, guava, pineapple and passion fruit dietary fibre concentrate. Food Chemistry. 2012;135(3):1520-1526.
- [42] Verkempinck SHE, Kyomugasho C, Salvia-Trujillo L, Denis S, Bourgeois M, Van Loey AM, Hendrickx ME, Grauwet T. Emulsion stabilizing properties of citrus pectin and its interactions with conventional emulsifiers in oil-in-water emulsions. Food Hydrocolloids. 2012;85(1):144-157.
- [43] Corradini SAS, Madrona GS, Visentainer JV, Bonafe EG, Carvalho CB, Roche PM, Prado IN. Sensorial and fatty acid profile of ice cream manufactured with milk of crossbred cows fed palm oil and coconut fat. Journal of Dairy Science. 2014;97(11):6745-6753.
- [44] Rabelo DM, Nascimento AL. Desenvolvimento e análise sensorial da geleia de polpa e casca de abacaxi com gengibre. Revista Academica Conecta FASF. 2014;3(1):1-14.
- [45] Lago-Vanzela ES, Ramin P, Umsza-Guez MA, Santos GV, Gomes E, Silva RD. Chemical and sensory characteristics of pulp and peel 'cajá-manga' (Spondias cytherea Sonn) jelly. Food Science and Technology. 2011;31(2):398-405.
- [46] Botelho L, Conceição A, Carvalho VD. Caracterização de fibras alimentares da casca e cilindro central do abacaxi 'smooth cayenne'. Ciência e Agrotecnologia. 2002:26(2):362-367.
- [47] Palmqvist E, Hahn-Hägerdal B. Fermentation of lignocellulosic hydrolysates. II: inhibitors and mechanisms of inhibition. Bioresource Technology. 2000;74(1):25-33.
- [48] Chitarra MIF, Chitarra AB. Pós-colheita de frutos e hortaliças: fisiologia e manuseio. 2nd ed. Lavras: UFLA; 2005.
- [49] Pardo MES, Cassellis MER, Escobedo RM, García EJ. Chemical characterisation of the industrial residues of the pineapple (Ananas comosus). Journal of Agricultural Chemistry and Environment. 2014;3(2):53-56.
- [50] Selvamuthukumaran M, Khanum F, Bawa AS. Development of sea buckthorn mixed fruit jelly. International Journal of Food Science & Technology. 2007;42(4):403-410.
- [51] Pinheiro ACM, Boas EVDBV, Lima LC. Influência do CaCl2 sobre a qualidade pós-colheita do abacaxi cv. Pérola. Food Science and Technology. 2005;25(1):32-36.
- [52] Ferreira MS, Santos MC, Moro TM, Basto GJ, Andrade RM, Gonçalves ÉC. Formulation and characterization of functional foods based on fruit and vegetable residue flour. Journal of Food Science and Technology. 2015;52(2):822-830.
- [53] Cao S, Yang Z, Zheng Y. Sugar metabolism in relation to chilling tolerance of loquat fruit. Food Chemistry. 2013;136(1):139-143.

- [54] Soares AS, Augusto PED, Júnior BRDCL, Nogueira CA, Vieira ÉNR, de Barros FAR, Stringheta PC, Ramos AM. Ultrasound assisted enzymatic hydrolysis of sucrose catalyzed by invertase: Investigation on substrate, enzyme and kinetics parameters. LWT-Food Science and Technology. 2019;107(1):164-170.
- [55] Joomwong A, Sornsrivichai J. Impact of cropping season in northern Thailand on the quality of smooth cayenne pineapple. II. Influence on physico-chemical attributes. International Journal of Agriculture and Biology. 2006;8(6):330-336.
- [56] Silva JM, Silva JP, Spoto MHF. Características físico-químicas de abacaxi submetido à tecnologia de radiação ionizante como método de conservação pós-colheita. Food Science and Technology. 2008;28(1):139-145.
- [57] Damodaran S, Parkin KL, Fennema OR. Química de alimentos de Fennema, 4nd ed. Porto Alegre: Artmed; 2010.
- [58] Gordon MH. The development of oxidative rancidity in foods. In: Pokorny J, Yanishlieva N, Gordon M. Antioxidant in Food: Practical Applications. Cambridge: Woodhead Publishing Limited; 2017.
- [59] Pettolino FA, Walsh C, Fincher GB, Bacic A. Determining the Polysaccharide Composition of Plant Cell Walls. Nature Protocols. 2012;7(9):1590-1607.
- [60] Rodriguez-Concepcion M, Avalos J, Bonet ML, Boronat A, Gomez-Gomez L, Hornero-Mendez D, Limon CM, Meléndez-Martínez AJ, Olmedilla-Alonso B, Palou A, Ribot J Rodrigo MJ, Zacarias L, Changfu Z. A global perspective on carotenoids: Metabolism, Biotechnology, and Benefits for Nutrition and Health. Progress in Lipid Research. 2018;70(1):62-93.
- [61] Damiani C, Almeida ACSD, Ferreira J, Asquieri ER, Boas V, Barros EV, Silva FAD. Doces de corte formulados com casca de manga. Pesquisa Agropecuária Tropical. 2011;41(3):360-369.
- [62] Uchoi D, Raju CV, Lakshmisha IP, Singh RR, Elavarasan, K. Antioxidative effect of pineapple peel extracts in refrigerated storage of Indian Mackerel. Fishery Technology. 2017;54(1):42-50.
- [63] Kongsuwan A, Suthiluk P, Theppakorn T, Srilaong V, Setha, S. Bioactive compounds and antioxidant capacities of phulae and nanglae pineapple. Asian Journal of Food and Agro-Industry. 2009;2(Special Issue): S44-S50.
- [64] Rodriguez-Amaya DB, Kimura M, Godoy HT, Amaya-Farfan J. Updated Brazilian database on food carotenoids: Factors affecting carotenoid composition. Journal of Food Composition and Analysis. 2008;21(6):445-463.
- [65] Grassin C, Coutel Y. Enzymes in fruit and vegetable processing and juice extraction. In: Whitehurst RJ, Van Oort M. Enzymes in Food Technology. 2nd ed. New Delhi: Blackwell Publishing Ltd; 2009.