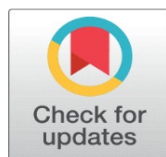


A COMPREHENSIVE REVIEW ON NEUROTOXICITY OF PYRETHROIDS

Zeeshan Ahmed ¹, Saman Athar ²

¹Project Assistant, Department of Microbiology, Central Drug Research Institute, Lucknow, Uttar Pradesh, India

²Ex-Student, Department of Zoology, Aligarh Muslim University, Aligarh, Uttar Pradesh, India



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Corresponding Author

Zeeshan Ahmed,
zeeshanfamily531995@gmail.com

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ABSTRACT

The natural pyrethrins produced by *Chrysanthemum cinerariaefolium* are converted into synthetic pyrethroids. Esters of chrysanthemum acid (ethyl, 2-dimethyl-3-(1-isobutenyl) cyclopropane-1-carboxylate) and halogenated derivatives of their acids and alcohols are included in them. Pyrethroids are frequently employed in menage illnesses and companion animal ectoparasite management solutions, but their infrequent usage in domestic settings raises concerns about exposure and unfavourable effects on people and more sophisticated animals. Post convinced exploration with a wide range of pyrethroids has indicated that the choreothetosis-expectoration (CS) pattern commonly appears as chemicals with the mode T-cyano-3-phenoxybenzylalcohol, such as deltamethrin, cypermethrin, and fenvalerate. General, extensively used bracket of Pyrethroid composites are determined grounded upon the symptomology of nonentity goods noted in neurophysiological tests. Numerous lines of evidence show that all pyrethroids and DDT analogues have a single major molecular target, the voltage-sensitive sodium channel. In biophysical and biochemical examinations, the changes in sodium channel functioning are nearly connected to the impact of these substances on complete neurons. The pyrethroid sodium channel discovery point demonstrates the strict stereo particularity anticipated by in vivo nonentity neurotoxicity estimates. Composites of type I and II exhibit qualitative improvements in sodium channel tail currents, divergent effects on entire neurons and in invertebrate muscle excitability. In order to determine whether this vast and significant collection of disorders forms a single "common medium" group or several groups for the purposes of cumulative problem assessment, knowledge of the molecular processes supporting pyrethroid neurotoxicity is immediately applicable.

Keywords: Pyrethroids, Neurotoxicity, Review

1. INTRODUCTION

India is a generally agricultural community. In India, new husbandry ways similar as increased irrigation, high yielding kinds, agrochemicals, and field mechanization have all helped to increase food product [Skolarczyk et al. \(2017\)](#), [Wylie et al. \(2016\)](#). Contemporaneously, the use of pesticides on a wide scale for mosquito, disease, and agrarian protection has expanded [Mehrotra \(1990\)](#), [Morgan \(1992\)](#), [Skolarczyk et al. \(2017\)](#). The riddle with pesticides is that they shield police, but their wide use frequently comes with pitfalls ranging from acute exposure to long term health consequences [Mehrotra \(1990\)](#), [Morgan \(1992\)](#), [Skolarczyk et al. \(2017\)](#).

Pesticides are divided into two orders: inorganic composites and organic composites. Pesticides similar as synthetic pyrethroids and organophosphates are generally used worldwide. The wide and expansive use of synthetic pyrethroids and organophosphates has rebounded in resistance, nonentity rejuvenescence, and health pitfalls [Morgan \(1992\)](#), [Toynton et al. \(2009\)](#).

Synthetic pyrethroids are created from the natural pyrethrins that *Chrysanthemum cinerariaefolium* produces. Halogenated derivatives of their acids and alcohols, as well as chrysanthemum acid esters, are included among them [Chrustek et al. \(2018\)](#), [Soderlund \(2012\)](#), [Costa \(2015\)](#), [Sethi et al. \(n.d.\)](#). Natural ingredients used in the *Chrysanthemum cinerariaefolium* extract are rapidly corrupted by light and have been substituted with synthetic derivatives that were first thought to be safe for humans and other sophisticated species. These contain 42 compounds and are classified as the fourth group of germicides by the WHO ([Chrustek et al., 2018](#)). The interaction of pyrethroids with sodium channels and the generation of sustained depolarization in neurons serve as their mechanisms of action. [Cárcamo et al. \(2017\)](#), [Chrustek et al. \(2018\)](#), [Soderlund \(2012\)](#), [Hussain \(n.d.\)](#), [Sethi et al. \(n.d.\)](#), [Wylie et al. \(2016\)](#).

Pyrethroids have been used to discourage pests and minimize crop losses for over a century [Spurlock and Lee \(1991\)](#). It has been set up that numerous known pyrethroids are dangerous to humans, mammals, marine creatures, and other healthy organisms [Frank et al. \(2018\)](#). Likewise, their prolonged presence in the ecosystem can affect in pollution, similar as impurity of groundwater, dislocation of agrarian product, and so on. Despite all sweats to minimize the negative impact of germicides on the terrain, there's still a pressing need to probe safer germicides [Ishaaya \(2003\)](#).

Pyrethroids mostly enter the body through skin contact, although they can also do so by inhalation or ingesting food or drink. [Hughes et al. \(2008\)](#), [Chrustek et al. \(2018\)](#), [Hughes and Edwards \(2010\)](#), [Hughes and Edwards \(2016\)](#), [Orsborne et al. \(2016\)](#), [Ranjesh et al. \(2013\)](#), [Singleton et al. \(2014\)](#). Professional job, water, nutrition, and household are the main exposure types [Chrustek et al. \(2018\)](#). It was established that the metabolites of pyrethroids were found in the urine following ingestion of semolina (pasta), rice, bread, morning cereals, and fruits from various locations. [Chrustek et al. \(2018\)](#), [Glorennec et al. \(2017\)](#), [Del Prado-Lu \(2015\)](#).

2. THE STATUS OF PYRETHROID INSECTICIDES CURRENTLY

Synthetic pyrethroid insecticides have been widely employed for more than three decades to reduce nonentity pests and disease vectors. Their use has increased to account for 18% of the market's total cash value by 2002 [Soderlund \(2012\)](#), [Pickett \(2004\)](#). Pyrethroids are effective against malaria and other microbiological diseases in addition to their use in pest management. The common use of pyrethroids in household germicides and companion animal ectoparasite control treatments, along with their restricted usage in the home environment, increases the risk of exposure and negative effects in the general populace. [Soderlund \(2012\)](#), [Naeher et al. \(2010\)](#), [Ostrea et al. \(2009\)](#), [Power and Sudakin \(2007\)](#). The Food Quality Protection Act (FQPA) of 1996 authorised nonsupervisory evaluation of the enrollments of pyrethroid germicide products in the United States. Since these categories of fungicides are thought to share a "common medium of toxicity," this legislation mandates that the United States Environmental Protection Agency take over the accretive danger evaluations for them.

When assessing whether this broad and significant class of germicides represents a single "common medium" group or numerous groups for the purposes of accretive threat assessment, knowledge of the molecular processes underlying pyrethroid neurotoxicity is immediately applicable. The default presumption of the reduced perceptivity of fetuses and newborns to the harmful effects of fungicides is a different nonsupervisory element added to the FQPA. By addressing this dereliction presupposition at the pharmacodynamic position, knowledge of the discriminational perceptivity of fetal, neonatal, and adult targets for the neurotoxic conduct of pyrethroids gives the power to improve the accuracy of threat assessments [Soderlund \(2012\)](#).

Marketable specifics, such as household fungicides, pet sprays, and cleansers, include nonentity control pyrethroids. Some pyrethroids are also used as nonentity repellents that may be applied to clothing and for direct lice treatments. The conflation analogues and derivations represent a diversified group over other strong germicides from the original pyrethrins. Although they're grounded on chemical changes that make them more dangerous and less environmentally degradable [The Pesticide Manual World Compendium. \(1997\)](#).

3. STRUCTURE AND INSECTICIDAL PROPERTIES OF PYRETROIDS

3.1. STRUCTURE AND CLASSES OF PYRETROIDS

Pyrethroids can be classified into two types on the basis of their mode of action [Table 1](#). Type I are based upon electrophysiological criteria. These compounds cause restlessness, in coordination and hypersensitivity followed by prostration and paralysis [Wylie et al. \(2016\)](#). Type II are classified on the basis of symptoms observed in pests, these compounds produce convulsive effects within minutes of dosing [Ray \(1991\)](#).

Pyrethrins, cinerins, and jasmolins are all contained in a common extract of natural pyrethrum from *Chrysanthemum cinerariaefolium*. Pyrethrin I is the most effective natural insecticide while pyrethrin II delivers more of the desired quick knock-down action against flying insects. The structure of pyrethrin I is an excellent starting point for addressing the molecular features required for insecticidal action, as well as how these features can be changed to maximise efficacy, reduce mammalian toxicity, and enhance stability. This will show how the guiding principles that contributed to the current range of synthetic pyrethroids were created. This will show how the guiding principles that contributed to the current range of synthetic pyrethroids were created [Elliott \(1971\)](#), [Elliott \(1976\)](#), [Sawicki, and Thain \(1962\)](#).

Table 1

Table 1 Classification of Synthetic Pyrethroids			
Pyrethrins	Type I Pyrethroids	Type II Pyrethroids	Reference
Constituents of natural pyrethrum extract	Derivatives of pyrethrins that do not include a cyano group and may elicit tumors	Derivatives of pyrethrins that include a cyano group and may elicit sinuous writhing and salivation	
Pyrethrin I	Allethrin	Cyfluthrin	(Sonia Sethi, n.d.)
Pyrethrin II	Bifenthrin	Deltamethrin	
Cinerin I	Permethrin	Cypermethrin	

Cinerin II	Pheothrin	Fenvalerate
Jasmolin I	Resemethrin	Fenpropathrin
Jasmolin II	Tefluthrin	Flucythrinate
	Tetramethrin	Flumethrin

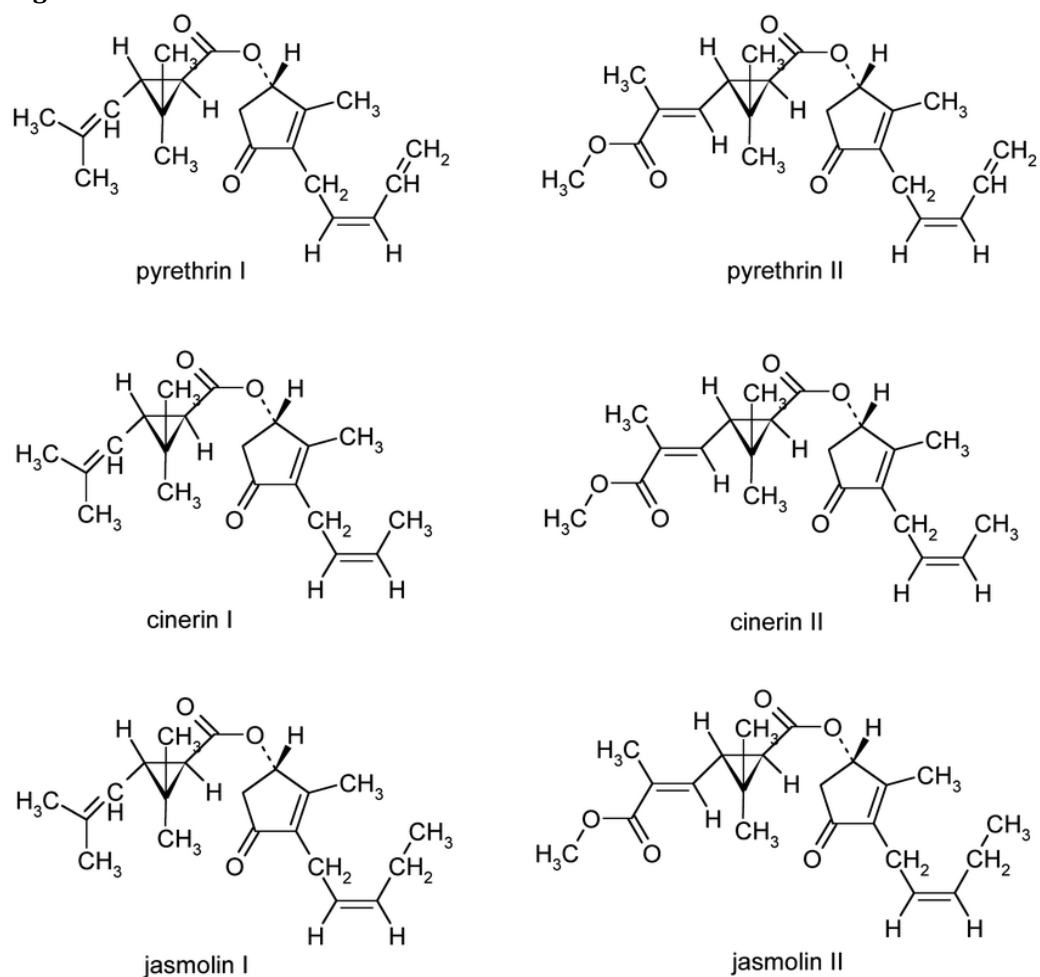
Figure 1**Figure 1** Structures of Natural Pyrethrins

Figure 2

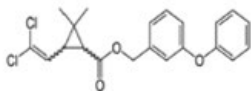
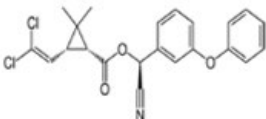
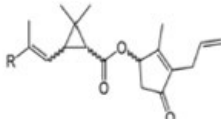
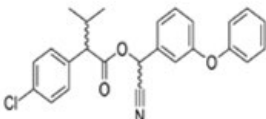
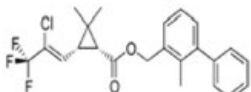
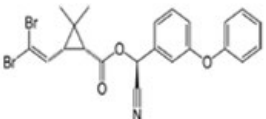
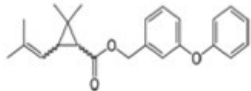
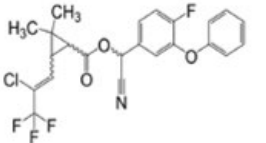
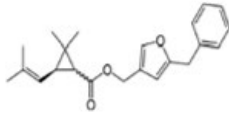
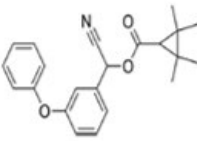
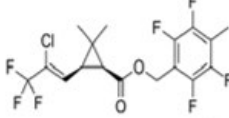
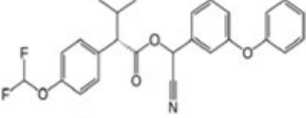
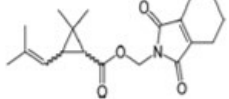
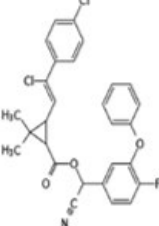
Type I	Molecule	Type II	Molecule
<u>Permethrin</u>		<u>Cypermethrin</u>	
Allethrin		Fenvalerate	
Bifenthrin		Deltamethrin	
Phenothrin		Cyfluthrin	
Resmethrin		Fenpropathrin	
Tefluthrin		Flucythrinate	
Tetramethrin		Flumethrin	

Figure 2 Pyrethroids Type I and Type II

Due to the many chiral carbons in cyclopropyl, the majority of pyrethroids have several stereoisomers. In addition, some pyrethroids have olefinic structures that are susceptible to cis or trans isomers. And because the activities or toxicity of different stereoisomers to different organisms are quite unequal, enrichment of active isomers can improve insecticidal activity while reducing toxicity to non-target organisms to some extent [Gerlach \(2012\)](#), [Nillos et al. \(2008\)](#), [Gan and Schlenk \(2008\)](#).

In the case of cyhalothrin, two chiral carbon atoms on the cyclopropane ring produce four optical isomers and two geometric isomers, yielding a total of eight isomers. There is also a chiral carbon atom in the cyanohydrin structure, which produces two optical isomers, R- and S-. As a result, there are sixteen optical isomers of cyhalothrin (eight pairs of optical enantiomers) [Lutnicka and K. A. \(2009\)](#).

3.2. SYNTHESIS OF PYRETHROIDS

To produce a concentrate comprising the six pyrethrin types—pyrethrin I, pyrethrin II, cinerin I, cinerin II, jasmolin I, and jasmolin II—organic detergents are used to uproot the active components of pyrethroids. The three rethrolones are pyrethrolone, jasmolone, and cinerolone. Pyrethrins that contain chrysanthemic acid are classified as type I, and those that contain pyrethric acid are classified as type II. Processing the flowers to grow the pyrethrin is a time-consuming process that varies by area.

The six esters that make up natural pyrethrins are composed of a monoterpenoid acid that has been partially conjugated to an oxylipin alcohol of the rethrolone class. Early feeding exploration suggests that the three rethrolones (pyrethrolone, jasmolone, and cinerolone) are derived from the octadecanoid pathway, while the two monoterpenoids (chrysanthemic acid and pyrethric acid) are derived from the plastidial 1-deoxy-D- xylulose-5-phosphate (DXP) terpenoid pathway [Barthel \(1961\)](#), [Goffinet and Locatelli \(1969\)](#), [Martel and Huynh \(1967\)](#), [Schechter et al. \(1949\)](#).

Figure 3

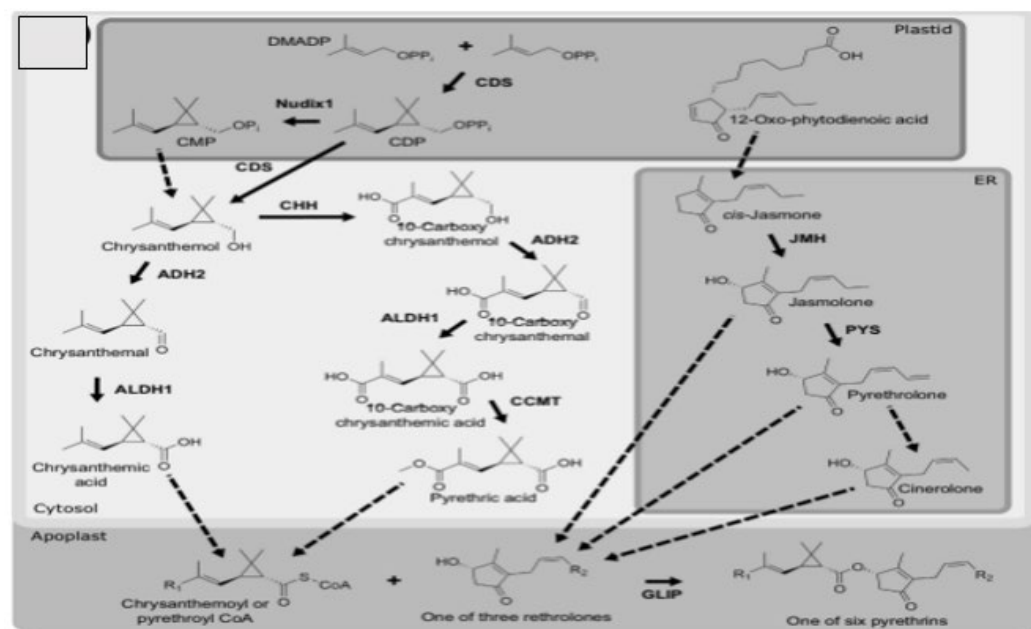


Figure 3 Synthesis of Natural Pyrethrins

The configurations of natural esters have been used to produce a variety of synthetic pyrethroid. Allethrin, constructed by Schechter, Green, and La Forge in 1949, was the first important synthetic emulsion that's still used moment. Natural esters pentadienyl side chains were condensed and simplified to make allethrin [Barthel \(1961\)](#), [Toynton et al. \(2009\)](#). Allethrolone, the alcoholic portion, is now commercially resolved, and the (+)- form, esterified with synthetic (+)- trans-chrysanthemic acid (+), yields S- bioallethrin, which is stereochemically original to pyrethrin I. Allethrin, bioallethrin, and S- bioallethrin are produced in lesser amounts than any other synthetic pyrethroid. Allethrin is used as a relief for natural pyrethrum when stocks are limited. Allethrin is frequently used in mosquito coils because of its advanced volatility and thermal adaptability than natural esters. 9 S-

Bioallethrin is a quick knockdown agent for certain insects, but it lacks the wide range of action of natural pyrethrins, including bioallethrin and allethrin [Elliott \(1971\)](#), [Rauch et al. \(1972\)](#). The coming synthetic pyrethroid to be commercially developed was Neopynamin (tetramethrin), which was published in 1964. Although neopynamin knocks insects down snappily when mixed into aerosols and analogous phrasings, the alcoholic portion isn't structurally related to other synthetic pyrethroids, and it isn't inescapably a strong payoff agent [Elliott \(1976\)](#), [Fales et al. \(1972\)](#), [Kato et al. \(1964\)](#). Presently, large-scale civilization of Dalmatian pyrethrum is demanded for artificial product of pyrethrin germicides. Before being collected, gutted, and ground up, pyrethrins build up in mature flower heads to a mass of one to two dry pounds [Chang et al. \(2014\)](#), [Sethi et al. \(n.d.\)](#). The powdered material can then be sold straight or combined with organic detergents to make insecticidal detergents and sprays [Kaneko \(2010\)](#). In addition to dry maquillages and liquid sprays used for small-scale ground-position care or large-scale upright operation compass of nonentity pests, pyrethrins are added to various accessories such as poultices and mosquito coils for specific nonentity safety. Wider spectrum germicides that work against a variety of non-entity pests include Coleoptera, Dyptera and Hemiptera (Homoptera and Heteroptera), Hymenoptera, Lepidoptera and Orthoptera, and Thysanoptera [Bradberry et al. \(2005\)](#), [Kaneko \(2010\)](#), [Kaneko \(2011\)](#), [Toynton et al. \(2009\)](#). They are scattered on food products before harvesting to control pests and also are used as germicides and protectives for homes. They are used on beast houses, in granges, in green houses and in veterinary drug considerably [Table 2](#). Pyrethroid effectiveness and selectivity are influenced by factors such as shape, significant structural features such as the ester and non-ester, specific chiral stereochemistry across the cyclopropane ring, physical parcels (for example, flying nonentity pests make good composites), and chemical parcels (e.g., more polar emulsion for knockdown). In order to boost the efficacy of germicides, the pyrethroids have been synthesised with compounds comparable to piperonyl butoxide, piperonyl sulfoxide, and sesamex. The pyrethroids used in commercial formulations contain a significant amount of additional, mostly toxic, inactive components [Costa \(2015\)](#).

Table 2

Table 2 Applications of Pyrethroids				
Pyrethroid	Insects	Crops	Other locations and applications	Reference
Allethrin	Flies, mosquitoes, ants	NIA	Tropical use in pet sprays and shampoos, residential, public health, animal housing	(Sonia Sethi, n.d.)
Bifenthrin	Beetles, weevil, houseflies, mosquitoes, lice, bedbugs, aphids, moths, cockroaches, locust	Alfalfa, haybeans, cantaloupes, cereals, cotton field and grassseed, hops, melons, watermelons.	NIA	
Bioresmethrin	Houseflies, mosquitoes, cockroaches	NIA	Household, public health, animal houses	
Cyfluthrin	Aphids, cabbage stem flea beetle, cockroaches, houseflies, mosquitoes,	Alfalfa, cereals, cotton, citrus, deciduous fruit, growing nuts, maize, oilseed,	Green houses	

	rape winter stem weevil	pears, potatoes, rice, sugarbeet, sugarcane, tobacco, vegetables.	
Cyhalothrin	Bedbugs, beetles, houseflies, lice, mosquitoes, moths, weevils	NIA	Public health, animal houses, inert surfaces
Cypermethrin	Cockroaches, flies, mosquitoes, moths	Cotton, lettuce, onions, pears, peaches, pecans, sugarbeets	Residential and commercial buildings, animal's houses
Deltamethrin	Aphids, beetles, bollworm, bud-worm, caterpillars, cicadas, totrix moths, weevils, whitefly, winter moths	Alfalfa, beet, cereals, coffee, cotton, figs, fruits, hops, maize, oilseed rape, olives, oil pahns, potatoes, rice, soybeans, sunflowers, tea, tobacco, vegetables.	Forests, households, animal houses, stored products
Esfenvalerate	Beetles, moths	Cabbage, cotton, fruit trees, grains, groundnuts, maize, potatoes, sorghum, soybeans, sugarcane, sunflowers, tomatoes, vegetables, wheat.	Ornamentals, non crop land.
Fenvalerate	Beetles, cockroaches, flies, locusts, mosquitoes, moths	Alfalfa hay, apples, beet, cereals, cotton, cucurbita, fruit, greenbeans, groundnuts, hops, maize, nuts, oilseed rape, olives, potatoes, sorghum, soybeans, squash, sugarcane, sunflower, vegetables, vines, tobacco.	Ornamentals, forestry, non-crop land.

3.3 MECHANISM OF ACTION IN INSECTS

Pesticides that have been introduced to cropland can be picked up by plants [Burridge and H. K. \(1997\)](#), eaten by animals, insects, or microorganisms in the soil, or travel downward in the soil [Lidova et al. \(2016\)](#) and either sticks to it or dissolves in water, or vaporises [Lutnicka and K. A. \(2009\)](#) and enters the atmosphere, or breaks down into less toxic compounds through microbial and chemical pathways or is leached out [Orsborne et al. \(2016\)](#), [Wang et al. \(2017\)](#). The stability and solubility properties of pesticides applied to soil play a big role in their destiny.

Pyrethrum compounds are inactivated and decomposed by exposure to light and air and are also broken down in water to nontoxic materials. Mild acids and alkalis degrade pyrethrins quickly as well [Burridge and H. K. \(1997\)](#), [Lidova et al. \(2016\)](#). Voltage-gated sodium channels are the primary target of pyrethroids neurotoxic effects on insects. Voltage-gated calcium and voltage-gated chloride channels are additional targets [Burridge and H. K. \(1997\)](#), [Lidova et al. \(2016\)](#), [Lutnicka and K. A. \(2009\)](#). Pyrethroids affect sodium ion channels in insect nervous systems, causing them to open faster, stimulating nerve cells and causing paralysis. Calcium signals control a number of neuronal growth pathways, while chloride channels regulate cell length, resting potential, and transepithelial transport [Orsborne et al. \(2016\)](#), [Wang et al. \(2017\)](#).

- 1) Type I pyrethroids:** Permethrin is a member of the first category of pyrethroids. This substance exists as a liquid as well as yellow-brown and brown crystals and is soluble in organic solvents [Toynton et al. \(2009\)](#). Permethrin comes in two optical stereoisomers; cis and trans. Studies show that cis-permethrin is more neurotoxic than trans-permethrin [Nasuti et al. \(2013\)](#). Additionally, permethrin nanometrics with hydrodispersive

characteristics are employed to stop the chemical from binding. Therefore, it may be promptly eliminated from the body because colloidal water dispersion prevents it from accumulating and reduces its period of retention in the body. [Davies et al. \(2007\)](#).

Skin, the digestive system, and the respiratory system are the three main routes through which the permethrin enters the body [Toynton et al. \(2009\)](#), [Wylie et al. \(2016\)](#). By accelerating impulse conduction, permethrin damages insect nerves, causing paralysis and eventual death. Voltage-gated sodium channels, such as Nav1.6, Nav1.3, and Nav1.8 in mammals and VGSCs in insects, are affected by the substance by causing them to open too early and delaying their inactivation [Soderlund \(2012\)](#), [Costa \(2015\)](#), [Sethi et al. \(n.d.\)](#). This pyrethroid operates on calcium channels by promoting calcium ion return transport, according to research to date. [Soderlund \(2012\)](#), [Costa \(2015\)](#), [Power and Sudakin \(2007\)](#).

- 2) **Type II pyrethroids:** Deltamethrin is a member of the first class of pyrethroids (Figure 1B). This medication is soluble in alcohol and acetone but lipophilic and insoluble in water. Colorless, white, and/or medium beige crystals have been discovered [Nillos et al. \(2008\)](#), [O'Reilly et al. \(2006\)](#). It is employed in farming. All of these pests are successfully controlled, including aphids, whiteflies, lice, tse-tse flies, fleas, ticks, spiders, ants, bees, bedbugs, and cockroaches. Mosquito nets are frequently treated with deltamethrin because it also protects against malaria vectors including *Aedes aegyptii* and *Anopheles gambiae* [Table 2 Gebreslassie et al. \(2012\)](#).

It is believed that the origin of deltamethrin's neurotoxic action is the prolonged activation of voltage-gated sodium channels, which results in neuronal membrane depolarization, repetitive discharges, synaptic abnormalities, and hyperexcitatory poisoning symptoms in insects. [Soderlund \(2012\)](#), [Costa \(2015\)](#). Deltamethrin also influences the function of calcium and chloride channels in neurons.

Pyrethroids are 2250 times more toxic to insects than higher mammals. Insects' smaller anatomy, more fragile sodium channels, and lower body temperatures account for this. [Sethi et al. \(n.d.\)](#)s. Pyrethroids have been shown to have a detrimental effect on the ion channels in neuronal membranes as well as the mitochondrial membranes of aquatic species including fish (carp and rainbow trout) and shellfish (crayfish, lobster). [Burridge and H. K. \(1997\)](#), [Cárcamo et al. \(2017\)](#), [Lidova et al. \(2016\)](#), [Lutnicka and K. A. \(2009\)](#), [Toynton et al. \(2009\)](#), [Wang et al. \(2017\)](#).

4. NEUROTOXICITY OF PYRETHROIDS

Neurotoxicity may be described as any unfavourable effects generated by chemical, biological or physical agents on the central or peripheral nervous system. Neurotoxicity has been demonstrated to have been associated with several substances, including metals (e.g. plum), industrial chemicals (e.g. acrylamide), solvents, natural poisons (e.g. domoic acid), pharmaceutical medicines (e.g. doxorubicin), misuse medicaments (e.g. ectasy), and pesticides [World Health Organization \(WHO\). \(2016\)](#). Because of some fundamental properties including aerobic metabolism reliance, existence of axonal transport and the neurotransmission process, the nervous system is particularly susceptible to injury [Singh et al. \(2012\)](#).

Moreover, the nervous system develops, which is believed to be more susceptible to neurotoxic chemicals in terms of replication, migration, differentiation, neuronal myelination, and synapse. Increased is that the blood-brain barrier is not fully established. In fact, some of the known neurotoxicants are predominantly developmental and quantitatively and qualitatively distinct symptoms of neurotoxicity in development and adult years (e.g., in case of lead or ethanol).

Neurotoxicants can be divided in four groups from a general mechanical perspective. These groupings are neuronopathic, axon-targeted and axonopathic, myelinopathic and neurotransmission affective. A series of chemicals can lead to toxicity that leads to loss of neurons by necrosis or by apoptosis (neuronopathy). This loss of neurons is irreversible and may lead to a loss of certain functions through a global encephalopathy or, if neuron subpopulations alone are affected [Singh et al. \(2012\)](#).

4.1. EFFECT ON NERVES

A crucial framework for examining molecular mechanisms of action is provided by the detrimental effects of pyrethroids on normal nerve function. Extracellular electrodes were used in early electrophysiological studies of pyrethrum's effects on nerve function to measure the compound nerve effect potential in pesticide- and pencil-prepared ventric nerve cords. The induction of recurrent releases, measured either as an increase in spontaneous activity or a range of potential for action generated by the one electric stimulation, which is followed by an electrical block, is one of the key characteristics of pyrethral and pyrethroid intoxication on the whole nerve level. These conclusions also include the fundamental reason for observing the effects of allethrin on nerve action potentials, especially modifies the transitory sodium conductance through the voltage-sensitive sodium channel. These findings were validated in a range of axonal preparations by subsequent research using numerous Type I Pyrethroids [Narahashi \(1962a\)](#), [Narahashi \(1962b\)](#), [Ruigt \(1985\)](#).

Pyrethroids have well-characterized effects on nerve axons and also affect other neuronal components. Both invertebrate and vertebrate sensory structures are extremely sensitive to pyrethroids and typically react differently than axonal preparations. The housefly larva sensor nerves, the cercal nerves of cockroach and the locust crural nerve sensory fibres create extremely extended spontaneously or ambiguously triggered high-frequency impulse trains. Two reaction patterns were found in comparison with the activity of a number of pyrethroid structures. Short sense explosions were related with Type I compounds in cockroach preparation (e.g. allethrin, tetramethrin), whereas locust chemicals categorised as Type II generated extended sensory explosions [Clements and May \(1977\)](#), [Gammon et al. \(1981\)](#), [Osborne and Hart \(1979\)](#).

Pyrethroids also influence synaptic transmission via the presynaptic nerve endpoint. Pyrethroid depolarization is carried out in motor terminals linked to insect larval wall body muscles, resulting in release of neurotransmitters. These effects are assessed as the first rise in the frequency of small postsynaptic excitatory potential in muscles, followed by neuromuscular blocking. Pyrethroids effects on the sodium channel led to nervous-terminal depolarization, as demonstrated by the fact that tetrodotoxin (TTX) prevented the sodium channel's pyrethroid-dependent function. [Salgado et al. \(1983\)](#).

In addition, the activities of the DDT must include any consideration of pyrethroid activity on nerves. Early physiological trials with DDT and pyrethroids

have shown that these structurally varied chemicals have had remarkably comparable effects on invertebrate axons and frog peripheral nerves. The later development of a number of hybrid DDT-pyrethroid insecticides that have structural components that may be exploited for insecticidal activities in both DDT and pyrethroid demonstrate another similar method of action for these two pesticide classes. [Holan et al. \(1978\)](#), [Narahashi \(1969\)](#), [Van Den Bercken et al. \(1979\)](#).

5. MECHANISM NEUROTOXICITY OF PYRETHROIDS

5.1. EFFECT ON INSECTS VOLTAGE GATED SODIUM CHANNELS

A pore generating sub-unit (sub-unit) plus an auxiliary sub-unit make up VGSCs [Dong \(2007\)](#). The component is divided into four domains (I-IV), each of which has six trans-membrane segments. The protein's amino and carboxy termini are found within the cell. The fifth and sixth transmembrane segments (S5 and S6), as well as the loop between them, which provides selectivity for Na⁺, together make up the channel pore [Dong \(2007\)](#). As a voltage sensor, the positively charged amino acids in S4 cause a conformational shift that causes the channel to open when the membrane depolarizes. Domains III and IV are connected by a cytoplasmic connection that acts as an inactivity gate. The two separate "gates" that regulate the four states in which VGSCs can reside are known as the activation gate and the inactivation gate. The channel is closed, and the door is open at the resting membrane potential. The channel opens when the membrane becomes depolarized, enabling Na⁺ to enter the cell.

The VGSC α subunit is encoded by a single gene in insects. This gene is known as para in *Drosophila melanogaster*. The mature transcripts from this gene's messenger RNA are alternatively divided into different combinations of exons. The transcripts go through an RNA editing process, which normally changes the base of the encoded amino acid in certain nucleotides. Alternate splitting and RNA editing are used to create VGSCs with unique gating properties. In *Drosophila melanogaster*, the auxiliary component TipE enhances the expression of the Para cell surface, heightens the Na⁺ current peak, and changes the kinetics of channel inactivation.

Users of a fire-polized glass micropipette can see the currents created by separately opening and shutting sodium channels in tiny pieces of a cell membrane by using patch clamping technology. [Hamill et al. \(1981\)](#). [1R,trans]-Tetramethrin created a population of changed sodium channels under patch cluster circumstances, in which sodium conductance and opening kinetics were unaffected but channel-open distribution durations were noticeably lengthened. [Zamponi et al. \(1997\)](#). In a preliminary communication, extension of one-channel currents by fenvalerate was also documented [Holloway et al. \(1984\)](#). The results of patch clamp studies contradict the pyrethroid's predicted effects on sodium channels and instead show that the pyrethroid only modifies sodium channel inactivation kinetics, with no appreciable impact on other characteristics.

Table 3

Table 3 List of some of the species with kdr mutations			
Mutation	Location in VGSC	Species	References
Leucine to Phenylalanine	DIIS6	Musca domestica, Blattella germanica, Plutella xylostella, Myzus persicae, Anopheles gambiae, Culex pipiens, Culex quinquefasciatus, Haematobia irritans, Leptinotarsa decemlineata, Ctenocephalides felis, Frankliniella occidentalis, Cydia pomonella.	Dong (2007)
Leucine to Serine	DIIS6	Culex pipiens, Anopheles gambiae	
Leucine to Histidine	DIIS6	Heliothis virescens	
Methionine to Threonine	DIIS4-S5 linker	Musca domestica, Haematobia irritans, Heliothis virescens	
Aspartate to Glycine	Amino terminus	Blattella germanica	
Glutamate to Lysine	DI and DII linker	Blattella germanica	
Cysteine to Arginine	DI and DII linker	Blattella germanica	
Proline to Leucine	Carboxy terminus	Blattella germanica	
Valine to Methionine	DIS6	Heliothis virescens	
Methionine to Isoleucine	DIIS1-S2 linker	Pediculus capitis	
Leucine to Phenylalanine	DIIS5	Pediculus capitis	
Threonine to Isoleucine	DIIS5	Plutella xylostella, Pediculus capitis	
Threonine to Cytosine	DIIS5	Frankliniella occidentalis	
Threonine to Valine	DIIS5	Ctenocephalides felis	
Phenylalanine to Isoleucine	DIIS6	Boophilus microplus	
Leucine to Proline	DIII-DIV linker	Varroa destructor	

There are genetic indications, in addition to electrophysiological data, showing pyrethroids target the VGSC. In the 1950s houseflies were initially documented in pyrethroid, or knockdown-resistant (KDR) [Busvine \(1951\)](#), [Davies et al. \(1958\)](#). The flying family's kdr and superkdr traits have shown that the VGSC Vssc1 housefly was directly responsible for their resistance to pyrethroids [Williamson et al. \(1993\)](#). Similar genetic mapping studies linked kdr and super-kdr traits in those species' VGSC genes in mosquitoes, German cockroaches, and tobacco budworms [Dong and Scott \(1994\)](#), [Severson et al. \(1997\)](#), [Taylor et al. \(1993\)](#). Two-point mutations were detected with Domain II by comparative sequencing analysis of Vssc1 kdr and super-kdr housefly with the wild-style gene [Williamson et al. \(1996\)](#). The first mutation, a change in phenylalanine in transmembrane segment 6, was seen in the two kdr and six super-kdr housefly strains. Only super-Kdr strains were found to have the second mutation, which changes methionine to threonine in the intracellular loop between transmembrane segments 4 and 5. In the sequence of VGSC genes from several species with kdr resistance, further changes [Table 3](#) have been found [Dong \(1997\)](#), [Dong \(2007\)](#), [Ingles et al. \(1996\)](#), [Miyazaki et al. \(1996\)](#), [Park et al. \(1997\)](#), [Soderlund et al. \(2008\)](#).

5.2. EFFECT ON GABA RECEPTOR- IONOPHORE COMPLEX

In recent years, intensive research has been done to ascertain whether or not the GABA receptor chloride ionophore complex of inhibitory synapses is a primary or secondary target for pyrethroids of type II. Although functional testing has indicated that the interaction of type II pyrethroids with the GABA mammalian receptor complex has been established in binding assays, this connection may not have much of a toxicogenic effect [Gammon et al. \(1982\)](#). Assessment of the effects of pyrethroid interactions with the chloride channel's TBPS bonding site is possible using chloride ion flow studies to test GABA receptor connections in mammalian brain preparations to their chloride canals [Bloomquist et al. \(1986\)](#), [Harris and Allan \(1985\)](#).

The most thorough analysis was used to compare the activities of the pyrethroid effects of deltamethrin, its benign enantiomer, and its insecticide noncyano counterpart, NRDC 157 [Bloomquist and Soderlund \(1985\)](#). GABA induced chloride absorption via deltamethrin inhibition into the mouse brain vesicles was inadequate and could not exceed 60% inhibition at 30/~ M.

The Type I ester NRDC 157 is predicted to be an inhibitor of chloride absorption by TBPS binding studies. Despite the likelihood that pyrethroid interactions with the TBPS site may alter the GABA receptorophore's ability to function, our results demonstrate that deltamethrin is 1000 times less efficient as a sodium channel activator than as an inhibitor of GABA-dependent chloride absorption. The incomplete stereospeutic properties of deltamethrin and its enantiomer inhibit chloride uptake are not compatible with the absolute stereospeutic character of both neurotoxicity and sodium channels, as determined by the results of intact nerves and the impact of these compounds [Ghiasuddin and Soderlund \(1985\)](#).

Pyrethroids only have a very strong effect on this target, according to functional tests with GABA-invertebrate species. Although the concentrations of deltamethrin affecting the GABA musculoskeletal receivers were several orders greater than those causing a deep disruption of the crayfish nerves through an effect on sodium channels that depended on voltage, the actions of GABA at the crayfish neuromuscular junction were antagonised by deltamethrin and other Type II pyrethroids. [Purves et al. \(2001\)](#).

5.3. PERIPHERAL TYPE BENZODIAZEPINE RECEPTORS

Pyrethroids lower the BBC Ro5-ability 4864's ability to bind to the "benzodiazepine receptor peripheral-type" site but have no effect on the binding of benzodiazepine radioligands to the benzodiazepine recognition site associated with the GABA receptor complex. [Gammon et al. \(1981\)](#), [Lawrence et al. \(1985\)](#). The capacity of type I and type II pyrethroids to behave as proconvulsants has recently been linked to their ability to interact with this site by lowering the threshold for the onset of pentylenetetrazole-induced seizures in rats. [Devaud and Murray \(1987\)](#), [Devaud et al. \(1986\)](#). Stereospecificity for neurotoxic isomers is suitable in the proconvulsing and receptor-binding effects both. It is important to acquire maximised in vivo provoking effects at dosages much below the levels necessary for pyrethroid-dependent acute poisoning [Devaud et al. \(1986\)](#). Additionally, Ro5-4864 was shown to delay the onset of detamethrine and permethrin in the cockroaches and to stop the GABA ligand flunitrazepam from binding to preparations of insect nerve and muscle receptors in mammals. The relevance of those findings as proof of pyrethroid interactions with either the GABA receptor complex or an insect-type

peripheral benzodiazepine receptor is uncertain because benzodiazepines are difficult to correlate with mammalian functions in insects [Abalis et al. \(1983\)](#), [Lummis and Sattelle \(1986\)](#).

5.4. NICOTINIC ACETYLCHOLINE RECEPTORS

Pyrethroid interactions with the nicotinic acetylcholine receptor were examined in the process of attaching the $[3H]$ perhydrohistrionico toxin (H2-HTX) to a locus connected to the acetylcholine gated ion channel. Pyrethrins, allethrin, resmethrin, and Tetramethrin were the most potent and rapid inhibitors of H2-HTX binding, but permethrin and other cyan-replaced esters were less potent and shown slow association. kinetics [Abbassy et al. \(1982\)](#). These pyrethroids also decreased Carbachol triggered $45Ca^{2+}$ intake, an acetylcholine analogue supposed to mimic flow via the relatively unspecified cholinergic receptor cation canal. However, after tests demonstrated that pyrethroids had no effects on $22Na^{+}$ absorption, which indicated that there was really a seeming impact on calcium absorption on calcium binding [Eldefrawi et al. \(1985\)](#). These findings suggest that the H2-HTX binding domain of the nicotinic acetylcholine receiver interacts with pyrethroids without affecting ion transport.

5.5. THE REGULATION OF CALCIUM AND ATP HYDROLYZING ENZYMES

The many Ca^{2+} ATPases are a group of ATPases involved in the action of pyrethroids and are thought to have a role in the stringent homeostatic regulation of calcium levels intracellular. Early research demonstrating the effects of DDT and pyrethroids on Ca^{2+} dependent ATPases as well as the causes for the control of calcium in pesticide action have all been covered elsewhere [Beeman \(1982\)](#). Pyrethroids inhibit two Ca^{2+} -ATPase activities in squid and cockroach nerve preparations: a Na^{+} - Ca^{2+} -ATPase that was supposed to be an ATP-module sodium-calcium tractor and was more sensitive to allethrin; and a Ca^{2+} - Mg^{2+} ATPase that was supposed to represent energy-dependent extrusion of calcium and was more sensitive to inhibition by type II pyrethroid [Clark and Matsumura \(1982\)](#).

Comparing the effectiveness of several pyrethroids as inhibitors in this test revealed that the Type II compounds were more potent than the Type I ones. The studies also showed that pyrethroids decreased the activity of adenylate cyclase when it was induced by calmodulin, indicating that interactions with calmodulin rather than ATPase or adenylate cyclase have acted as a mediator for all activities in this system. Additionally, DDT and pyrethroids lessen the activation of phosphodiesterase by calmodulin [Rashatwar and Matsumura \(1985\)](#).

6. CONCLUSION

For at least two centuries, the insecticide capabilities of the pyrethrins, the natural insecticides present in pyrethrum flowers and other extracts, were identified. A variety of synthetic analogues were produced and tested between 1940 and 1970 by pyrethrins (named pyrethroids). Some of the compounds have showed an outstanding insecticide activity, including allethrin, tetramethrin, and resmethrin and have been produced as commercial home, storage, and veterinary insecticides. However, they all had one thing in common: natural pyrethroid esters are unstable in the environment, which prevents them from being widely employed in agricultural production. The global agricultural chemicals industry reaffirmed its

interest in this kind of material, prompting Permethrin, the first photostable pyrethroid, to conduct extensive research and development. A number of previous pyrethroids and other kinds of insecticides have been superseded by the various photostable pyrethroids that were produced as a consequence of these efforts, and they have proven to be incredibly efficient farm insecticides. As a result, pyrethroids have a comprehensive identification of one of the four or five main classes of synthetic insecticides.

Various lines of evidence suggest the major location of pyrethroid activity in the neurological system. In insects, pyrethroids quickly cause indications of poisoning, indicating action on the neurological system (lack of coordinated movement, convulsive activities phases and final paralytic). Post induced research with a wide range of pyrethroids has indicated that the choreoathetosis-salivation (CS) syndrome often occurs as substances like deltamethrin, cypermethrin, and fenvalerate, which have the mode T-cyano-3-phenoxybenzylalcohol. General, widely used classification of Pyrethroid compounds (a structurally various group producing a syndrome) and compounds of the Type II compounds (predominately Cyan-3-phenoxybenzyl esters, producing a CS syndrome), are determined based upon the symptomology of insect effects noted in neurophysiological tests.

However, all these activities may not be implicated in disrupting nerve activity, pyrethroid insects interact with a range of neurochemical procedures. The voltage-sensitive sodium channel is the main molecular target for all pyrethroids and DDT analogues in both insects, according to a number of lines of evidence. Biophysical and biochemical investigations have shown that the effects of these medicines on intact neurons are closely tied to changes in sodium channel activity. The pyrethroid sodium channel detection site demonstrates the strict stereo specificity expected by in vivo insect neurotoxicity estimates. Type I and type II chemicals differ in their qualitative effects on intact neurons, sodium channel tail currents, and the excitability of the muscles in the vertebrate skeleton. Additionally, sodium channel kinetics has considerable effect on the Type I/Type H categorization method. The diverse poisoning symptoms seen in insects appear to be sufficiently explained by the multiple sensory and motor nerve pathways of these qualitatively variable effects on the sodium channel.

An in vitro test, the impacts of pyrethroids on additional neurochemical targets seems not to be significant in vivo for these chemicals neurotoxin effects. Although pyrethroids Type II interactions with the GABA receptor complex's ionophore chloride component can inhibit GABA-dependent chloride flow, the quantities required to affect this system are several magnitude levels higher than those that would disrupt sodium channel function. At addition, pyrethroid activities in the GABA receptor do not show the strict stereo specificity indicated by the acute neurotoxicity measurements.

The link between pyrethroids and the nicotinic acetylcholine receptor does not seem to affect their functioning, unlike the activity of pyrethroids in a GABA receptor complex. This result is in accordance with the failure of physiological studies to identify the effects of pyrethroids on post-synaptic receptors, and it shows that these interactions with the nicotinic acetylcholine receptor are not harmful. Finally, there are murky connections between the neurotoxicity of DDT and pyrethroids and their inhibitory effects on ATPases and calmodulin. DDT and pyrethroid effects on mitochondrial Na⁺, K⁺, and Mg²⁺ ATPase have been identified, although these effects do not fully explain their effects on the nerves as a whole. Although regulating intraneuronal calcium concentration is more directly related to nerve function,

pyrethroids may not directly account for those drugs' effects on neuronal excitability by altering calcium homeostasis.

CONFLICT OF INTERESTS

None.

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REFERENCES

- Abalis, I. M., Eldefrawi, M. E., and Eldefrawi, A. T. (1983). Biochemical Identification of Putative GABA/Benzodiazepine Receptors In House Flythorax Muscles. *Pesticide Biochemistry and Physiology*, 20(1), 39-48. [https://doi.org/10.1016/0048-3575\(83\)90119-0](https://doi.org/10.1016/0048-3575(83)90119-0).
- Abbassy, M. A., Eldefrawi, M. E., and Eldefrawi, A. T. (1982). Allethrin Interactions With The Nicotinic Acetylcholine Receptor Channel. *Life Sciences*, 31(15), 1547-1552. [https://doi.org/10.1016/0024-3205\(82\)90045-5](https://doi.org/10.1016/0024-3205(82)90045-5).
- Barthel, W. F. (1961). Synthetic Pyrethroids. *Advan. Pest Control Res*, 4, 33.
- Beeman, R. W. (1982). Recent Advances In Mode of Action of Insecticides. *Annual Review of Entomology*, 27, 253-281. <https://doi.org/10.1146/annurev.en.27.010182.001345>.
- Bloomquist, J. R., Adams, P. M., and Soderlund, D. M. (1986). Inhibition of 3-Aminobutyriacc Id-Stimulated Chloride Flux In Mousbe Rain Vesicle By Polychlorocycloalkane And Pyrethroid Insecticides. *Neurotoxicology*, 7(3), 11-20.
- Bloomquist, J. R., and Soderlund, D. M. (1985). Neurotoxic Insecticides Inhibit GABA-Dependent Chloride Uptake by Mouse Brain Vesicles. *Biochemical and Biophysical Research Communications*, 133(1), 37-43. [https://doi.org/10.1016/0006-291x\(85\)91838-8](https://doi.org/10.1016/0006-291x(85)91838-8).
- Bradberry, S. M., Cage, S. A., Proudfoot, A. T., and V. J. A. (2005). Poisoning Due To Pyrethroids. *Toxicology Research*, 24, 93-106. <https://doi.org/10.2165/00139709-200524020-00003>.
- Burridge, L. E., and H. K. (1997). Lethality of Pyrethrins to Larvae and Postlarvae of The American Lobster (*Homarus Americanus*). *Ecotoxicology and Environment Safety*, 38, 150-154. <https://doi.org/10.1006/eesa.1997.1571>
- Busvine, J. R. (1951). Mechanism of resistance to insecticide in houseflies. *Nature*, 168(4266), 193-195. <https://doi.org/10.1038/168193a0>.
- Chang, F., Dutta, S., Becnel, J. J., Estep, A. S., and Mascal, M. (2014). Synthesis Of The Insecticide Prothrin and Its Analogues from Biomass-Derived 5-(Chloromethyl) Furfural. *Journal of Agricultural and Food Chemistry*, 62(2), 476-480. <https://doi.org/10.1021/jf4045843>.
- Chrustek, A., Hołyńska-Iwan, I., Dziembowska, I., Bogusiewicz, J., Wróblewski, M., Cwynar, A., and Olszewska-Słonina, D. (2018). Current Research on The Safety of Pyrethroids Used as Insecticides. *Medicina*, 54(4). <https://doi.org/10.3390/medicina54040061>.
- Clark, J. M., and Matsumura, F. (1982). Two Different Types of Inhibitory Effects of Pyrethroids on Nerve Ca- and Ca + Mg-Atpase Activity In The Squid, *Loligo Pealei*. *Pesticide Biochemistry and Physiology*, 18(2), 180-190. [https://doi.org/10.1016/0048-3575\(82\)90104-3](https://doi.org/10.1016/0048-3575(82)90104-3).

- Clark, J. M., and Matsumura, F. (1987). The Action of Two Classes of Pyrethroids on The Inhibition of Brain Na-Ca and Ca + Mg ATP Hydrolyzing Activities of The American Cockroach. *Comp. Biochemistry and Physiology*, 86, 135-145. [https://doi.org/10.1016/0742-8413\(87\)90156-3](https://doi.org/10.1016/0742-8413(87)90156-3).
- Clements, A. N., and May, T. E. (1977). The Actions of Pyrethroids Upon The Peripheral Nervous System and Associated Organs In The Locust. *Pesticide Science*, 8(6), 661-680. <https://doi.org/10.1002/ps.2780080611>.
- Costa, L. G. (2015). The Neurotoxicity of Organochlorine and Pyrethroid Pesticides. *Handbook of Clinical Neurology*, 131, 135-148. <https://doi.org/10.1016/B978-0-444-62627-1.00009-3>.
- Cárcamo, J. G., Aguilar, M. N., Carreño, C. F., Vera, T., Arias-Darraz, L., Figueroa, J. E., Romero, A. P., Alvarez, M., and Yañez, A. J. (2017). Consecutive Emamectin Benzoate and Deltamethrin Treatments Affect The Expressions and Activities of Detoxification Enzymes in the Rainbow Trout (*Oncorhynchus Mykiss*). *Comparative Biochemistry and Physiology. Toxicology And Pharmacology*, 191, 129-137. <https://doi.org/10.1016/j.cbpc.2016.10.004>.
- Davies, M., Keiding, J., and Von Hosten, C. G. (1958). Resistance To Pyrethrins and To Pyrethrinspiperonyl Butoxide in a Wild Strain of *Musca Domestica* L. In Sweden. *Nature*, 182(4652), 1816-1817. <https://doi.org/10.1038/1821816a0>.
- Davies, T. G. E., Field, L. M., Usherwood, P. N. R., and Williamson, M. S. (2007). DDT, Pyrethrins, Pyrethroids and Insect Sodium Channels. *IUBMB Life*, 59(3), 151-162. <https://doi.org/10.1080/15216540701352042>.
- Del Prado-Lu, J. L. (2015). Insecticide residues in soil, water, and eggplant fruits and farmers' health effects due to exposure to pesticides. *Environmental Health and Preventive Medicine*, 20(1), 53-62. <https://doi.org/10.1007/s12199-014-0425-3>.
- Devaud, L. L., Szot, P., And Murray, T. F. (1986). PK 11195 Antagonism of Pyrethroid-Induced Proconvulsant Activity. *European Journal of Pharmacology*, 121(2), 269-273. [https://doi.org/10.1016/0014-2999\(86\)90499-1](https://doi.org/10.1016/0014-2999(86)90499-1).
- Devaud, L., and Murray, T. F. (1987). Interactions of Pyrethroid Insecticides With The Peripheral-Type Benzodiazepine Receptor. *Social Neuroscience [Abstr.]*, 13, 1230.
- Dong, K. (1997). A Single Amino Acid Change In The Para Sodium Channel Protein Is Associated With Knockdown-Resistance (Kdr) To Pyrethroid Insecticides In German Cockroach. *Insect Biochemistry and Molecular Biology*, 27(2), 93-100. [https://doi.org/10.1016/S0965-1748\(96\)00082-3](https://doi.org/10.1016/S0965-1748(96)00082-3).
- Dong, K. (2007). Insect Sodium Channels and Insecticide Resistance. *Invertebrate Neuroscience*, 7(1), 17-30. <https://doi.org/10.1007/s10158-006-0036-9>.
- Dong, K., and Scott, J. G. (1994). Linkage of Kdr-Type Resistance and the Para-Homologous Sodium Channel Gene In German Cockroaches (*Blattella Germanica*). *Insect Biochemistry And Molecular Biology*, 24(7), 647-654. [https://doi.org/10.1016/0965-1748\(94\)90051-5](https://doi.org/10.1016/0965-1748(94)90051-5).
- Eldefrawi, M. E., Sherby, S. M., Abalis, I. M., and Eldefrawi, A. T. (1985). Interactions of Pyrethroid and Cyclodiene Insecticides With Nicotinic Acetylcholine and GABA Receptors. *Neurotoxicology*, 6(2), 47-62.
- Elliott, M. (1971). The Relationship Between The Structure and the Activity of Pyrethroids. *Bulletin of the World Health Organization*, 44(1-3), 315-324.
- Elliott, M. (1976). Properties and Applications of Pyrethroids. *Environmental Health Perspectives*, 14, 3-13. <https://doi.org/10.2307/3428357>.
- Fales, J. H. et al. (1972). Relative Effectiveness of Pyrethroid Insecticides. *Soap, Cosmetics, Chemical Specialties*, 48, 60.

- Frank, D. F., Miller, G. W., Harvey, D. J., Brander, S. M., Geist, J., Connon, R. E., and Lein, P. J. (2018). Bifenthrin Causes Transcriptomic Alterations In Mtor and Ryanodine Receptor-Dependent Signaling and Delayed Hyperactivity In Developing Zebrafish (Danio Rerio). *Aquatic Toxicology*, 200, 50-61. <https://doi.org/10.1016/j.aquatox.2018.04.003>.
- Gammon, D. W., Brown, M. A., And Casida, J. E. (1981). Two Classes of Pyrethroid Action In the Cockroach. *Biochern. Pesticide Biochemistry and Physiology*, 15(2), 181-191. [https://doi.org/10.1016/0048-3575\(81\)90084-5](https://doi.org/10.1016/0048-3575(81)90084-5).
- Gammon, D. W., Lawrence, L. J., and Casida, J. E. (1982). Pyrethroid Toxicology: Protective Effects of Diazepam and Phenobarbital In The Mouse and the Cockroach. *Toxicology and Applied Pharmacology*, 66(2), 290-296. [https://doi.org/10.1016/0041-008x\(82\)90294-0](https://doi.org/10.1016/0041-008x(82)90294-0).
- Gebreslassie, B. H., Yao, Y., and You, F. (2012). Design Under Uncertainty of Hydrocarbon Biorefinery Supply Chains: Multiobjective Stochastic Programming Models, Decomposition Algorithm, and A Comparison Between Cva R and Downside Risk. *Aiche Journal*, 58(7), 2155-2179. <https://doi.org/10.1002/aic.13844>.
- Gerlach, R. W. (2012). Chiral Chemistry and Toxicity Assessments for Pyrethroid Pesticides. *Journal of the American Chemical Society*. <https://doi.org/10.1021/bk-2012-1099.ch002>.
- Ghiasuddin, S. M., and Soderlund, D. M. (1985). Pyrethroid Insecticides, 24, 200-206. [https://doi.org/10.1016/0048-3575\(85\)90129-4](https://doi.org/10.1016/0048-3575(85)90129-4).
- Glorennec, P., Serrano, T., Fravallo, M., Warembourg, C., Monfort, C., Cordier, S., Viel, J. F., Le Gléau, F., Le Bot, B., and Chevrier, C. (2017). Determinants of Children's Exposure To Pyrethroid Insecticides In Western France. *Environment International*, 104, 76-82. <https://doi.org/10.1016/j.envint.2017.04.007>.
- Goffinet, B., and Locatelli, A. (1969). Separation of Dtrans-Chrysanthemic Acid From Its Optical and Geometrical Isomers. *Chem [Abstr.]*. Fr Pat. 1,536,458, 71, 90923w.
- Hamill, O. P., Marty, A., Neher, E., Sakmann, B., and Sigworth, F. J. (1981). Improved Patch-Clamp Techniques for High Resolution Current Recording From Ceils and Cell-Free Membrane Patches. *Pflügers Archiv - European Journal of Physiology*, 391(2), 85-100. <https://doi.org/10.1007/BF00656997>.
- Harris, R. A., and Allan, A. M. (1985). Functional Coupling of 3,-Aminobutyric Acid Receptors to Chloride Channels In Brain Membranes. *Science*, 228(4703), 1108-1110. <https://doi.org/10.1126/science.2581319>.
- Holan, G., O'Keefe, D. F., Virgona, C., and Walser, R. (1978). Structural and Biological Link Between Pyrethroids and DDT In New Insecticides. *Nature*, 272(5655), 734-736. <https://doi.org/10.1038/272734a0>.
- Holloway, S. F., Salgado, V. L., Wu, C. H., and Narahashi, T. (1984). Maintained Opening of Single Na Channels By Fenvalerate. *Social Neuroscience [Abstr.]*, 10, 864.
- Hughes, E. A., Flores, A. P., Ramos, L. M., Zalts, A., Richard Glass, C., and Montserrat, J. M. (2008). Potential Dermal Exposure To Deltamethrin and Risk Assessment For Manual Sprayers: Influence of Crop Type. *Science of the Total Environment*, 391(1), 34-40. <https://doi.org/10.1016/j.scitotenv.2007.09.034>.
- Hughes, M. F., and Edwards, B. C. (2010). In Vitro Dermal Absorption of Pyrethroid Pesticides In Human and Rat Skin. *Toxicology and Applied Pharmacology*, 246(1-2), 29-37. <https://doi.org/10.1016/j.taap.2010.04.003>.

- Hughes, M. F., and Edwards, B. C. (2016). In Vivo Dermal Absorption of Pyrethroid Pesticides In the Rat. *Journal of Toxicology and Environmental Health. Part A*, 79(2), 83-91. <https://doi.org/10.1080/15287394.2015.1109571>.
- Hussain, M. (n.d.). *Environmental Degradation : Realities and Remedies/Mumtaz Hussain (1st Ed)*.
- Ingles, P. J., Adams, P. M., Knipple, D. C., and Soderlund, D. M. (1996). Characterization of Voltage-Sensitive Sodium Channel Gene Coding Sequences from Insecticide-Susceptible And Knockdown Resistant House Fly Strains. *Insect Biochemistry and Molecular Biology*, 26(4), 319-326. [https://doi.org/10.1016/0965-1748\(95\)00093-3](https://doi.org/10.1016/0965-1748(95)00093-3).
- Ishaaya, I. (2003). Introduction: Biorational Insecticides-Mechanism and Application. *Archives of Insect Biochemistry And Physiology*, 54(4), 144. <https://doi.org/10.1002/arch.10111>.
- Pickett, J. A., (2004). New Opportunities In Neuroscience, But A Great Danger That Some May Be Lost. In D. J. Beadle, I. R. Mellor and P. N. R. Usherwood (Eds.), *Neurotox 2003: Neurotoxicological Targets From Functional Genomics And Proteomics* . Society Of Chemical Industry. 1-10.
- Kaneko, H. (2010). Pyrethroid Chemistry and Metabolism. In. In R. Krieger (Ed.), *Hayyes' Handbook of Pesticide Toxicology (3rd Ed)*. <https://doi.org/10.1016/B978-0-12-374367-1.00076-8>.
- Kaneko, H. (2011). Pyrethroids: Mammalian Metabolism and Toxicity. *Journal of Agricultural and Food Chemistry*, 59(7), 2786-2791. <https://doi.org/10.1021/jf102567z>.
- Kato, T., Ueda, K., and Fujimoto, K. (1964). New Insecticidally Active Chrysanthemates. *Agricultural and Biological Chemistry*, 28(12), 914-915. <https://doi.org/10.1080/00021369.1964.10858319>.
- Lawrence, L. J., Gee, K. W., and Yamamura, H. I. (1985). Interactions of Pyrethroid Insecticides With Chloride Ionophore-Associated Binding Sites. *Neurotoxicology*, 6(2), 87-98.
- Lidova, J., Stara, A., Kouba, A., and V. J. (2016). The Effects of Cypermethrin on Oxidative Stress and Antioxidant Biomarkers In Marbled Crayfish (*Procambarus Fallax F. Virginalis*). *Neuro Endocrinology Letters*, 37(Suppl1), 53-59.
- Lummis, S. C. R., and Sattelle, D. B. (1986). Binding sites for [3H]GABA[3, H]flunitrazepam and [35S]TBPS in insect CNS. *Neurochemistry International*, 9(2), 287-293. [https://doi.org/10.1016/0197-0186\(86\)90065-3](https://doi.org/10.1016/0197-0186(86)90065-3).
- Lutnicka, H., and K. A. (2009). Pyrethroids as A Predisposing Factor In Fish Diseases. *Ochr. Środ. Zasobów Natl*, 41, 285-292.
- Martel, J., and Huynh, C. (1967). Synthèse de l'acide chrysanthémique II. *Bull. Soc. Chem.*, 985.
- Mehrotra, K. N. (1990). Pyrethroids Resistant Insect Pest Management. *Pesticide Research Journal*, 2(1), 44-52.
- Miyazaki, M., Ohyama, K., Dunlap, D. Y., and Matsumura, F. (1996). Cloning and Sequencing of The Paratype Sodium Channel Gene from Susceptible and Kdr-Resistant German Cockroaches (*Blattella Germanica*) and House Fly (*Musca Domestica*). *Molecular and General Genetics*, 252(1-2), 61-68. <https://doi.org/10.1007/s004389670007>.
- Morgan, M. D. R. (1992). *The BMA Guide To Pesticides, Chemicals and Health*. Title. Published on Behalf of The British Medical Association By Edward Arnold.
- Morgan, M. K. (2012). *Children's Exposures to Pyrethroid Insecticides at Home: A Review of Data Collected In Published Exposure Measurement Studies*

- Conducted In The United States. *International Journal of Environmental Research and Public Health*, 9(8), 2964e2985. <https://doi.org/10.3390/ijerph9082964>.
- Naeher, L. P., Tulve, N. S., Egeghy, P. P., Barr, D. B., Adetona, O., Fortmann, R. C., Needham, L. L., Bozeman, E., Hilliard, A., and Sheldon, L. S. (2010). Organophosphorus and Pyrethroid Insecticide Urinary Metabolite Concentrations In Young Children Living In A Southeastern United States City. *Science of the Total Environment*, 408(5), 1145-1153. <https://doi.org/10.1016/j.scitotenv.2009.10.022>.
- Narahashi, T. (1962a). Effect of The Insecticide Allethrin on Membrane Potentials of Cockroach Giant Axons. *Journal of Cellular and Comparative Physiology*, 59, 61-65. <https://doi.org/10.1002/jcp.1030590108>.
- Narahashi, T. (1962b). Nature of The Negative After-Potential Increased By The Insecticide Allethrin in Cockroach Giant Axons. *Journal of Cellular and Comparative Physiology*, 59, 67-76. <https://doi.org/10.1002/jcp.1030590109>.
- Narahashi, T. (1969). Mode of Action of Ddta Nd Allethrin On Nerve: Cellular And Molecular Mechanisms. *Residue Reviews*, 25, 275-288. https://doi.org/10.1007/978-1-4615-8443-8_21.
- Nasuti, C., Carloni, M., Fedeli, D., Gabbianelli, R., Di Stefano, A., Serafina, C. L., Silva, I., Domingues, V., and Ciccocioppo, R. (2013). Effects of Early Life Permethrin Exposure on Spatial Working Memory and on Monoamine Levels In Different Brain Areas of Pre-Senescent Rats. *Toxicology*, 303, 162-168. <https://doi.org/10.1016/j.tox.2012.09.016>.
- Nillos, M. G., Gan, J., and Schlenk, D. (2008). Chemical Analysis and Enantioselective Toxicity of Pyrethroids. *Journal of The American Chemical Society*, 991, 400-414. <https://doi.org/10.1021/bk-2008-0991.ch018>.
- O'Reilly, A. O., Khambay, B. P., Williamson, M. S., Field, L. M., Wallace, B. A., and Davies, T. G. (2006). Modelling Insecticide-Binding Sites In The Voltage-Gated Sodium Channel. *Biochemical Journal*, 396(2), 255-263. <https://doi.org/10.1042/BJ20051925>.
- Orsborne, J., Deraedt Banks, S., Hendy, A., Gezan, S. A., Kaur, H., Wilder-Smith, A., Lindsay, S. W., and Logan, J. G. (2016). Personal Protection of Permethrin-Treated Clothing Against Aedes Aegypti, The Vector of Dengue And Zika Virus, In The Laboratory. *PLOS ONE*, 11(5), E0152805. <https://doi.org/10.1371/journal.pone.0152805>.
- Osborne, M. P., and Hart, R. J. (1979). Neurophysiological Studies of The Effects of Permethrin Upon Pyrethroid Resistant (Kdr) and Susceptible Strains of Dipteran Larvae. *Pesticide Science*, 10(5), 407-413. <https://doi.org/10.1002/ps.2780100507>.
- Ostrea, E. M., Jr., Bielawski, D. M., Posecion, N. C., Jr., Corrión, M., Villanueva-Uy, E., Bernardo, R. C., Jin, Y., Janisse, J. J., and Ager, J. W. (2009). Combined Analysis of Prenatal (Maternal Hair and Blood) and Neonatal (Infant Hair, Cord Blood And Meconium) Matrices To Detect Fetal Exposure of Environmental Pesticides. *Environmental Research*, 109(1), 116-122. <https://doi.org/10.1016/j.envres.2008.09.004>.
- Park, Y., Taylor, M. F., and Feyereisen, R. (1997). A Valine 421 to Methionine Mutation In IS6 of the Hscp Voltage-Gated Sodium Channel Associated With Pyrethroid Resistance In Heliothis Virescens F. *Biochemical and Biophysical Research Communications*, 239(3), 688-691. <https://doi.org/10.1006/bbrc.1997.7511>.

- Power, L. E., and Sudakin, D. L. (2007). Pyrethrin and Pyrethroid Exposures In The United States: A Longitudinal Analysis of Incidents Reported to Poison Centers. *Journal of Medical Toxicology*, 3(3), 94-99. <https://doi.org/10.1007/BF03160917>.
- Purves, D., Augustine, G. J., Fitzpatrick, D., Hall, W. C., LaMantia, A.-S., Mooney, R. D., Platt, M. L., and L. E. W. (2001). *Neuroscience* (2nd editio). Oxford University Press.
- Ranjakesh, M. R., Naghili, B., Goldust, M., and Rezaee, E. (2013). The Efficacy of Permethrin 5% Vs. Oral Ivermectin For the Treatment of Scabies. *Annals of Parasitology*, 59(4), 189-194.
- Rashatwar, S. S., and Matsumura, F. (1985). Interaction of DDT and Pyrethroids With Calmodulin and Its Significance In The Expression of Enzyme Activities Of Phosphodiesterase. *Biochemical Pharmacology*, 34(10), 1689-1694. [https://doi.org/10.1016/0006-2952\(85\)90635-5](https://doi.org/10.1016/0006-2952(85)90635-5).
- Rauch, F., Lhoste, J., and Birg, M. L. (1972). Proprieties Insecticides Du D-Trans Chrysanthemate De D-Allethrolone. *Mededel. Fak. Landbouw. Wetenschap. Gent*, 37, 755.
- Ray, D. E. (1991). Pesticides derived from plants and other organisms. Academic Press. (International Association of Bridge, Structural, Ornamental and Reinforcing Iron Workers J. H. *Journal of Engineering Research L. (Eds.). (Ed.)*).
- Ruigt, G. S. F. (1985). Pyrethroids. In *Comprehensive Insect Physiology* G. A. Kerkut and L. I. Gilbert (Eds.), *Biochemistry and pharmacology*, 12, 183-262.
- Salgado, V. L., Irving, S. N., and Miller, T. A. (1983). The Importance of Nerve Terminal Depolarization In Pyrethroid Poisoning of Insects. *Pesticide Biochemistry and Physiology*, 20(2), 169-182. [https://doi.org/10.1016/0048-3575\(83\)90021-4](https://doi.org/10.1016/0048-3575(83)90021-4).
- Sawicki, R. M., and Thain, E. M. (1962). Insecticidal Activity of Pyrethrum Extract And Its Four Insecticidal Constituents Against House Flies. IV. Knock-Down Activities of The Four Constituents. *Journal of The Science of Food And Agriculture*, 13(5), 292-297. <https://doi.org/10.1002/jsfa.2740130504>.
- Schechter, M. S., Green, N., and La Forge, F. B. (1949). Constituents of Pyrethrum Flowers. *Journal of The American Chemical Society*, XXIV Cinerolone and The Synthesis of Related Cyclopentenolones, 71, 3165. <https://doi.org/10.1021/ja01177a065>.
- Sethi, S., Mathur, N., and P. B. (n.d.). Synthetic pyrethroids: A review. *International Journal of Scientific and Engineering Research*, January, 20(1).
- Severson, D. W., Anthony, N. M., Andreev, O., and Ffrench-Constant, R. H. (1997). Molecular Mapping of Insecticide Resistance Genes In The Yellow Fever Mosquito (*Aedes Aegypti*). *Journal of Heredity*, 88(6), 520-524. <https://doi.org/10.1093/oxfordjournals.jhered.a023148>.
- Singh, A. K., Tiwari, M. N., Prakash, O. S.M., and Singh, M. P. (2012). A Current Review of Cypermethrin-Induced Neurotoxicity and Nigrostriatal Dopaminergic Neurodegeneration. *Current Neuropharmacology*, 10(1), 64-71. <https://doi.org/10.2174/157015912799362779>.
- Singleton, S. T., Lein, P. J., Farahat, F. M., Farahat, T., Bonner, M. R., Knaak, J. B., and Olson, J. R. (2014). Characterization of A-Cypermethrin Exposure In Egyptian Agricultural Workers. *International Journal of Hygiene And Environmental Health*, 217(4-5), 538-545. <https://doi.org/10.1016/j.ijheh.2013.10.003>.

- Skolarczyk, J., Pekar, J., and Nieradko-Iwanicka, B.. (2017). Immune Disorders Induced by Exposure to Pyrethroid Insecticides. *Postepy Hig Med Dosw (Online)*, 71(0)(I), 446-453. <https://doi.org/10.5604/01.3001.0010.3827>
- Soderlund, D. M. (2008). Pyrethroids, Knockdown Resistance and Sodium Channels. *Pest Management Science*, 64(6), 610-616. <https://doi.org/10.1002/ps.1574>.
- Soderlund, D. M. (2012). Molecular Mechanisms of Pyrethroid Insecticide Neurotoxicity: Recent Advances. *Archives of Toxicology*, 86(2), 165-181. <https://doi.org/10.1007/s00204-011-0726-x>.
- Spurlock, F., and Lee, M. (2008). Synthetic Pyrethroid Use Patterns, Properties, and Environmental Effects. *ACS (Am. Chem. Soc.) Symp. [Ser.]*. <https://doi.org/10.1021/bk-2008-0991.ch001>.
- Taylor, M. F., Heckel, D. G., Brown, T. M., Kreitman, M. E., and Black, B. (1993). Linkage of Pyrethroid Insecticide Resistance To a Sodium Channel Locus In The Tobacco Budworm. *Insect Biochemistry and Molecular Biology*, 23(7), 763-775. [https://doi.org/10.1016/0965-1748\(93\)90064-y](https://doi.org/10.1016/0965-1748(93)90064-y).
- The Pesticide Manual World Compendium. (1997). C. D. S. Tomlin (Ed.).
- Toynton, K., Luukinen, B., Buhl, K., and S. D. (2009). Permethrin technical fact sheet. National Pesticide Information Center.
- Wang, Y., Lv, L., Yu, Y., Yang, G., Xu, Z., Wang, Q., and Cai, L. (2017). Single and Joint Toxic Effects of Five Selected Pesticides on The Early Life Stages of Zebrafish (Denio Renio). *Chemosphere*, 170, 61-67. <https://doi.org/10.1016/j.chemosphere.2016.12.025>.
- Williamson, M. S., Denholm, I., Bell, C. A., and Devonshire, A. L. (1993). Knockdown Resistance (Kdr) To DDT and Pyrethroid Insecticides Maps To A Sodium Channel Gene Locus In The Housefly (Musca Domestica). *Molecular and General Genetics*, 240(1), 17-22. <https://doi.org/10.1007/BF00276878>.
- Williamson, M. S., Martinez-Torres, D., Hick, C. A., And Devonshire, A. L. (1996). Identification of Mutations In The Housefly Para-Type Sodium Channel Gene Associated With Knockdown Resistance (Kdr) To Pyrethroid Insecticides. *Molecular and General Genetics*, 252(1-2), 51-60. <https://doi.org/10.1007/BF02173204>.
- World Health Organization (WHO). (2016). Pesticide Evaluation Scheme, Vector Ecology and Management. World Health Organization.
- Wylie, B. J., Hauptman, M., Woolf, A. D., and Goldman, R. H. (2016). Insect Repellants During Pregnancy In The Era of The Zika Virus. *Obstetrics and Gynecology*, 128(5), 1111-1115. <https://doi.org/10.1097/AOG.0000000000001685>.
- Zamponi, G. W., Bourinet, E., Nelson, D., Nargeot, J., and Snutch, T. P. (1997). Crosstalk Between G Proteins and Protein Kinase C Mediated By The Calcium Channel A(1) Subunit. *Nature*, 385(6615), 442-446. <https://doi.org/10.1038/385442a0>.
- Van Den Bercken, J., Kroese, A. B. A., and Akkermans, L. M. A. (1979). Effects of Insecticides on The Sensory Nervous System. In *Neurotoxicology of Insecticides and Phermones*, 210, 183-210. https://doi.org/10.1007/978-1-4684-0970-3_10.